

RepeatExplorer pipeline



What is RepeatExplorer ?

Implementation of principles described in:

- Repetitive DNA in the pea (*Pisum sativum* L.) genome: comprehensive characterization using 454 sequencing and comparison to soybean and *Medicago truncatula* (BMC Genomics 2007, 8:427)
- Graph-based clustering and characterization of repetitive sequences in next-generation sequencing data (BMC Bioinformatics 2010, 11:378)
- TAREAN: a computational tool for identification and characterization of satellite DNA from unassembled short reads. Nucleic Acids Res., doi:10.1093/nar/gkx257(2017)

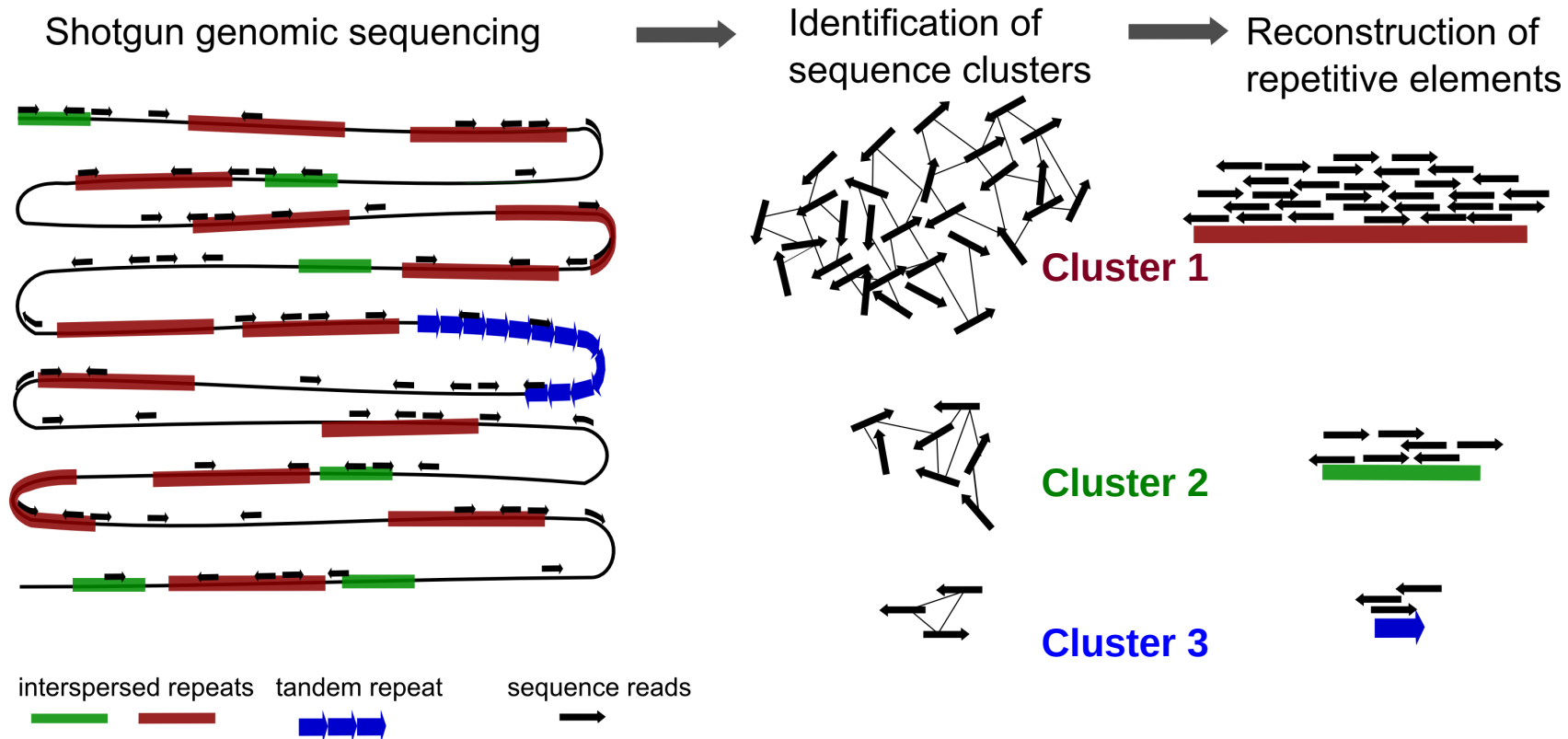
Protocols

- Global analysis of repetitive DNA from unassembled sequence reads using RepeatExplorer2. Nature Protocols 15:3745–3776.

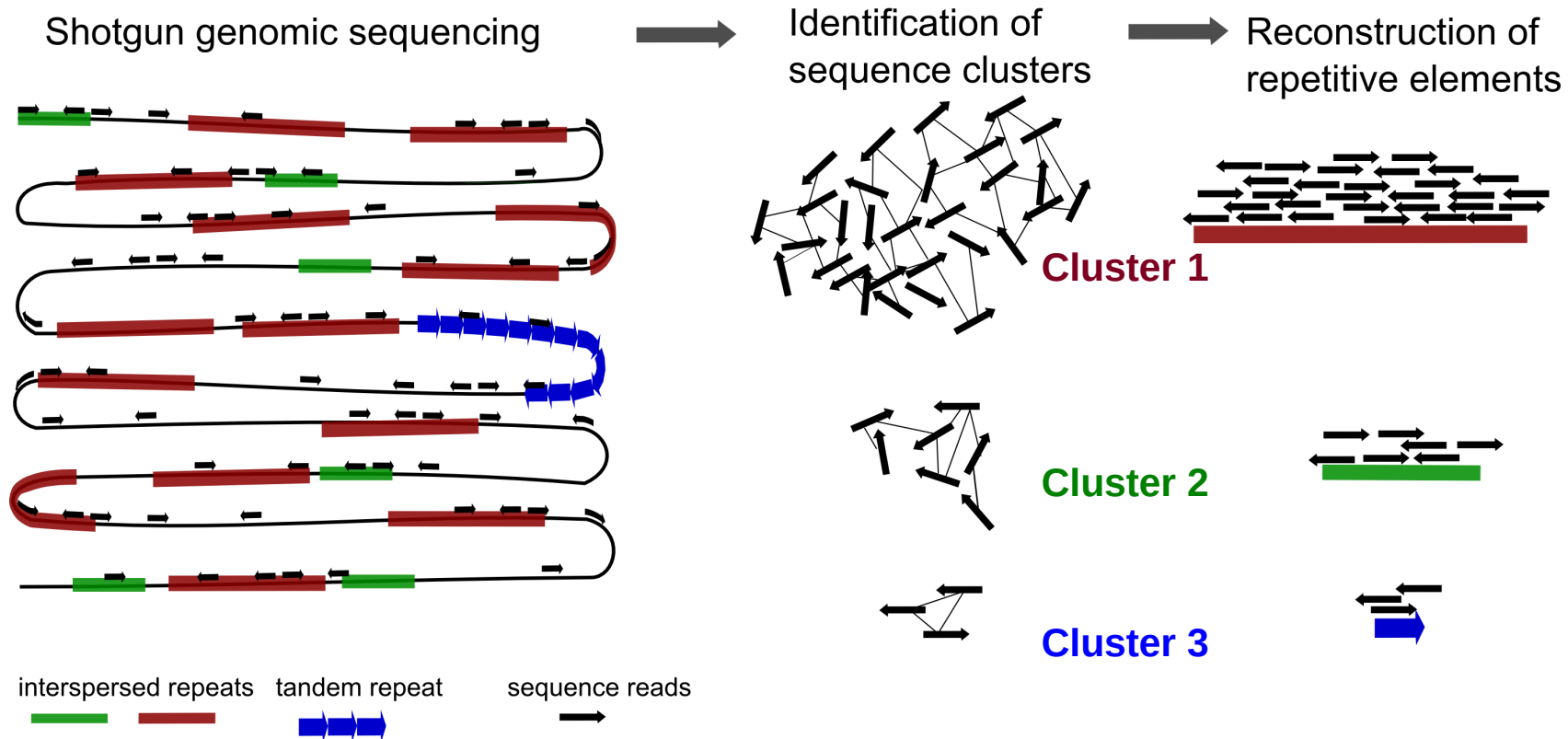
Available Tools:

- NGS data preprocessing
- RepeatExplorer2 pipeline
- TAREAN pipeline
- Chip-Seq analysis
- Domain based ANnotation of Transposable Element – DANTE
- Profrep
- Visualization

Principle of RepeatExplorer

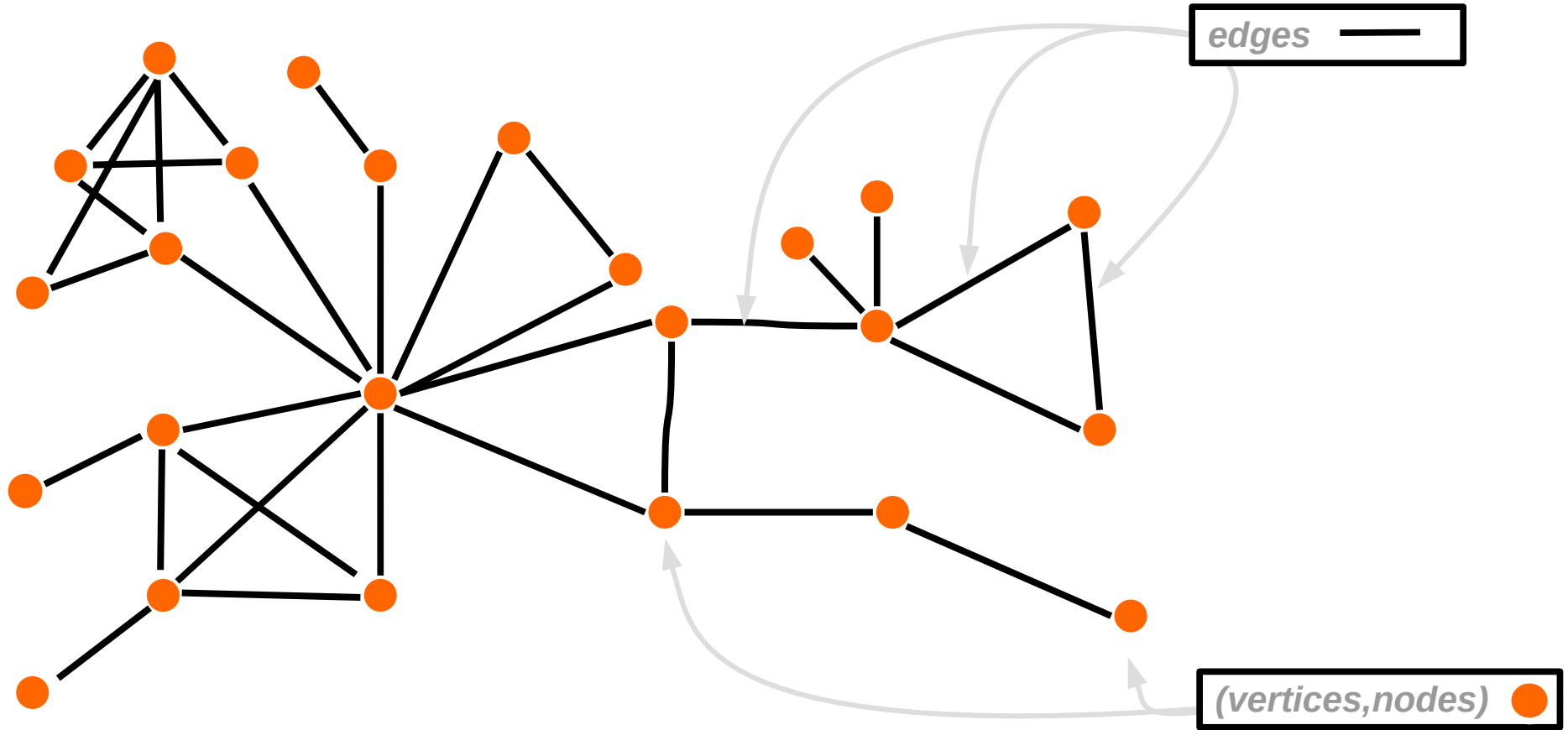


Principle of RepeatExplorer

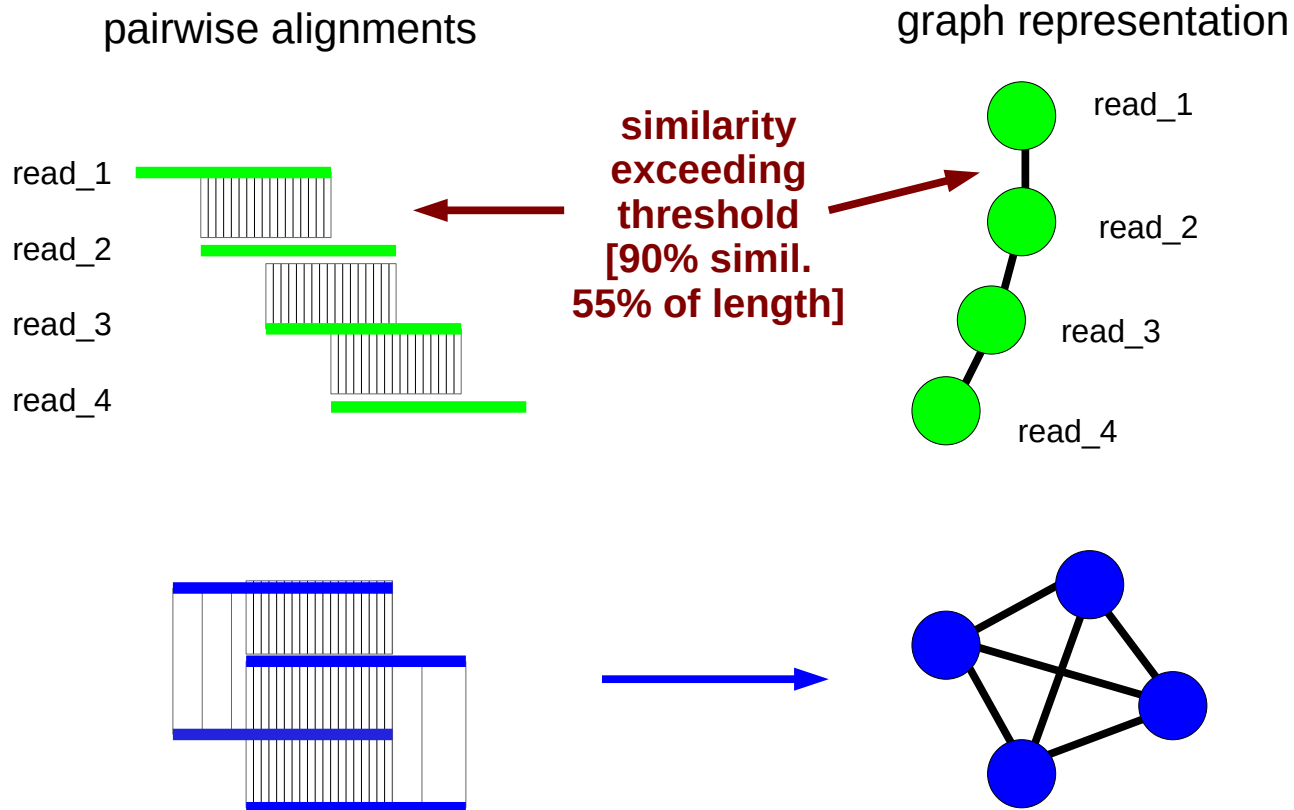


CLUSTER = a set of frequently overlapping reads = REPEAT FAMILY

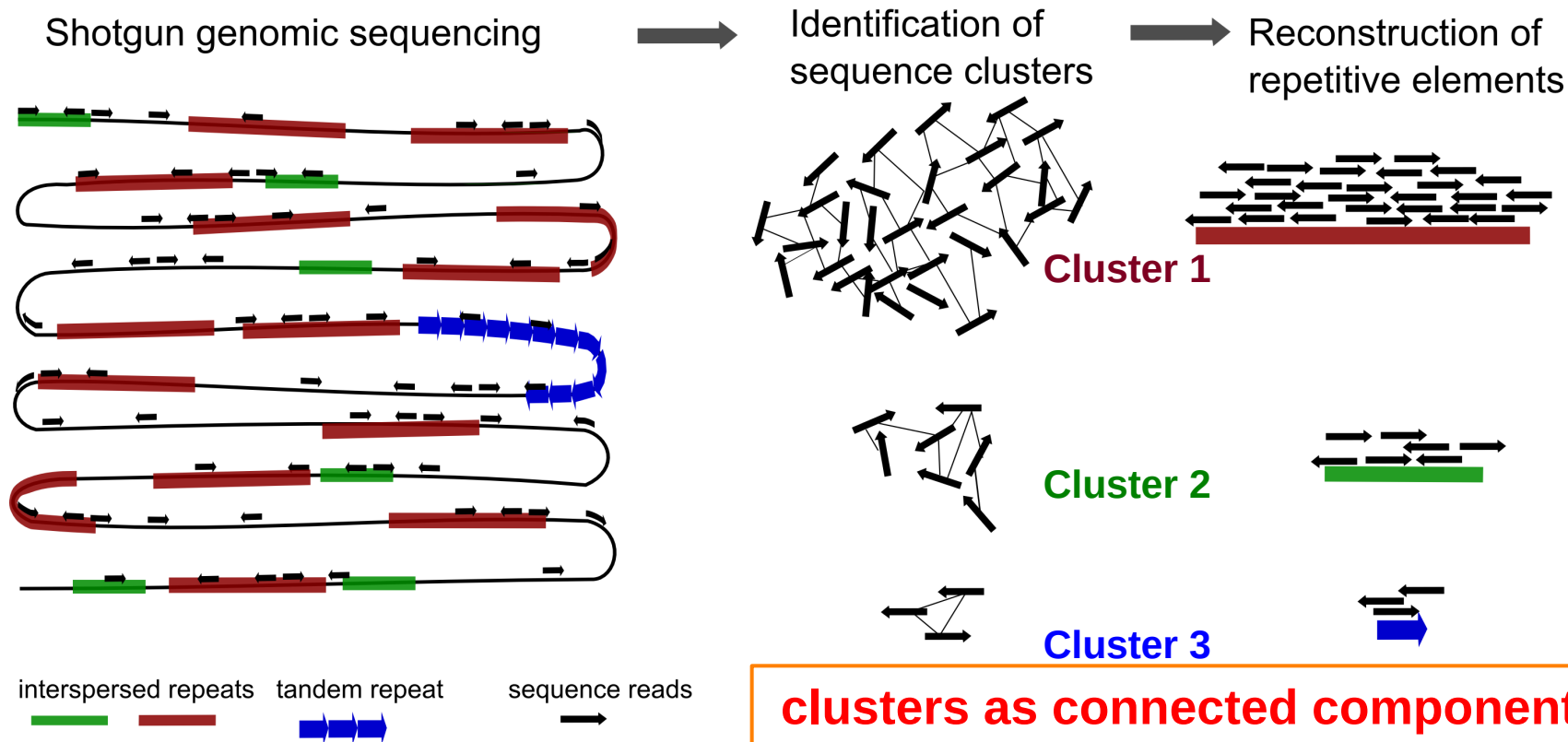
Graph Based Representation of Sequence Reads



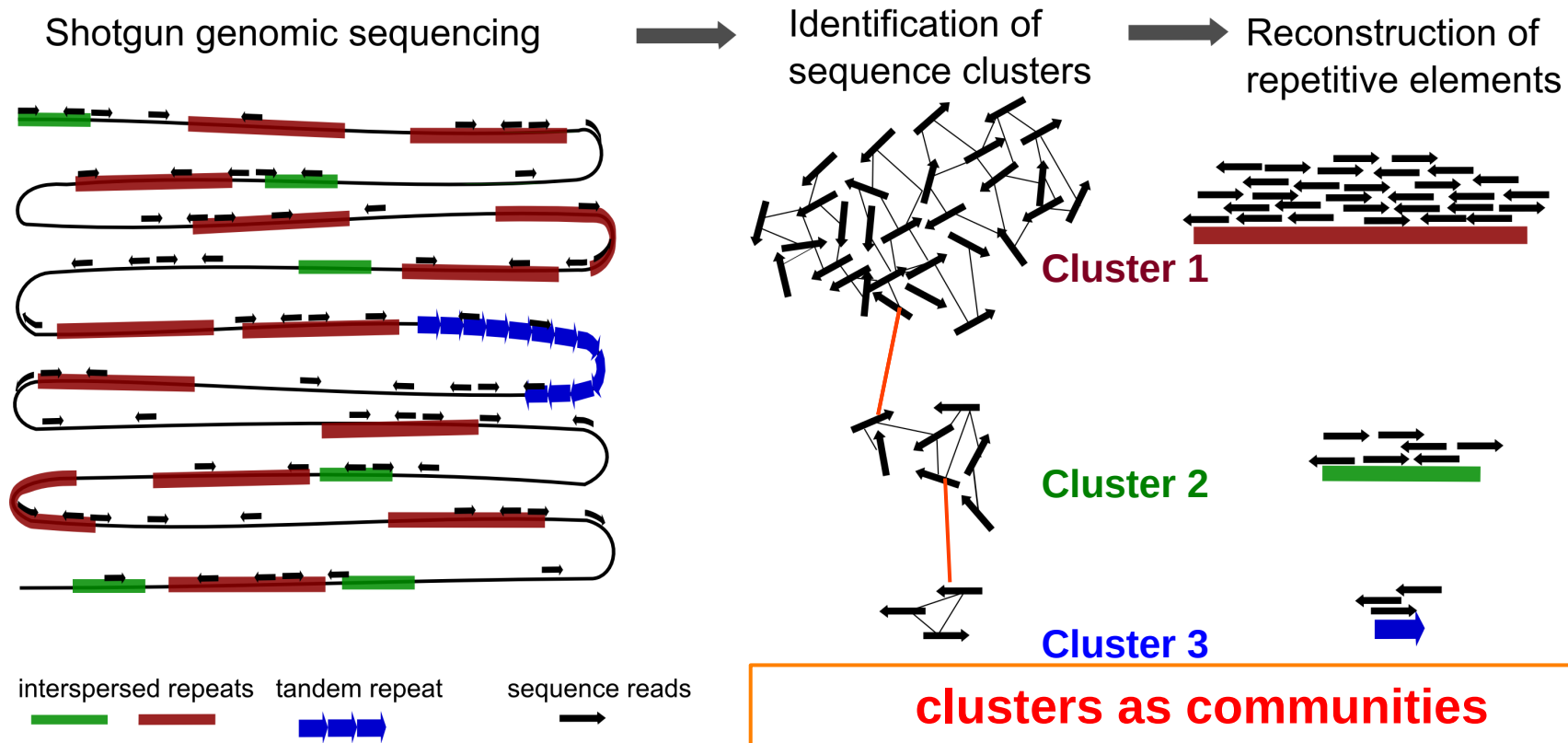
Graph Based Representation of Sequence Reads



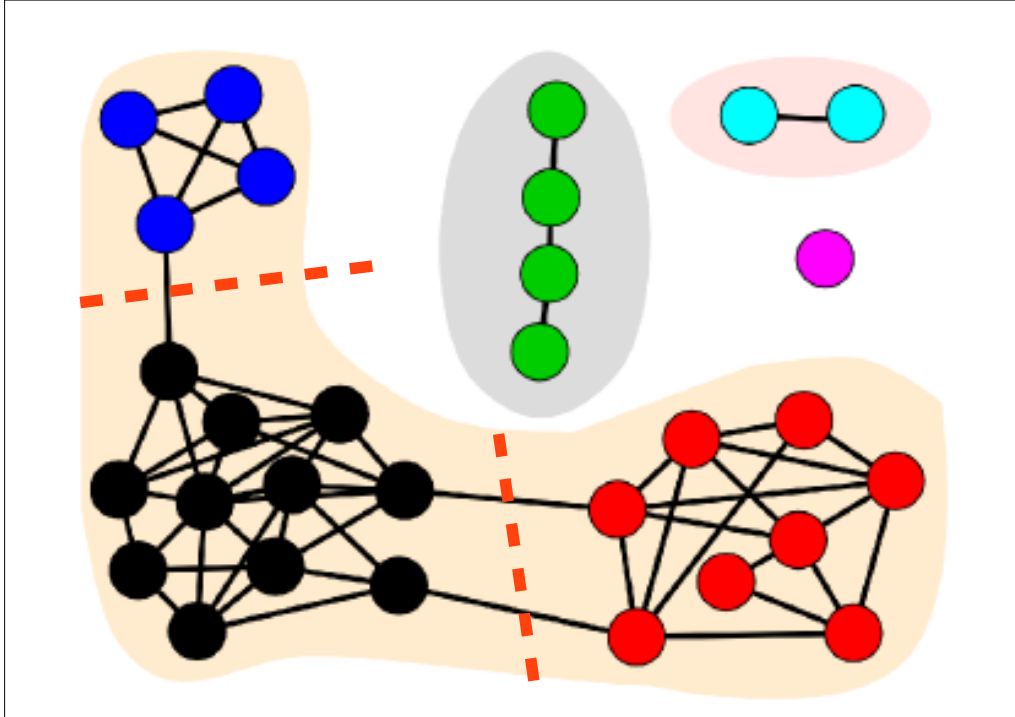
Principle of RepeatExplorer



Principle of RepeatExplorer



Graph Based Clustering

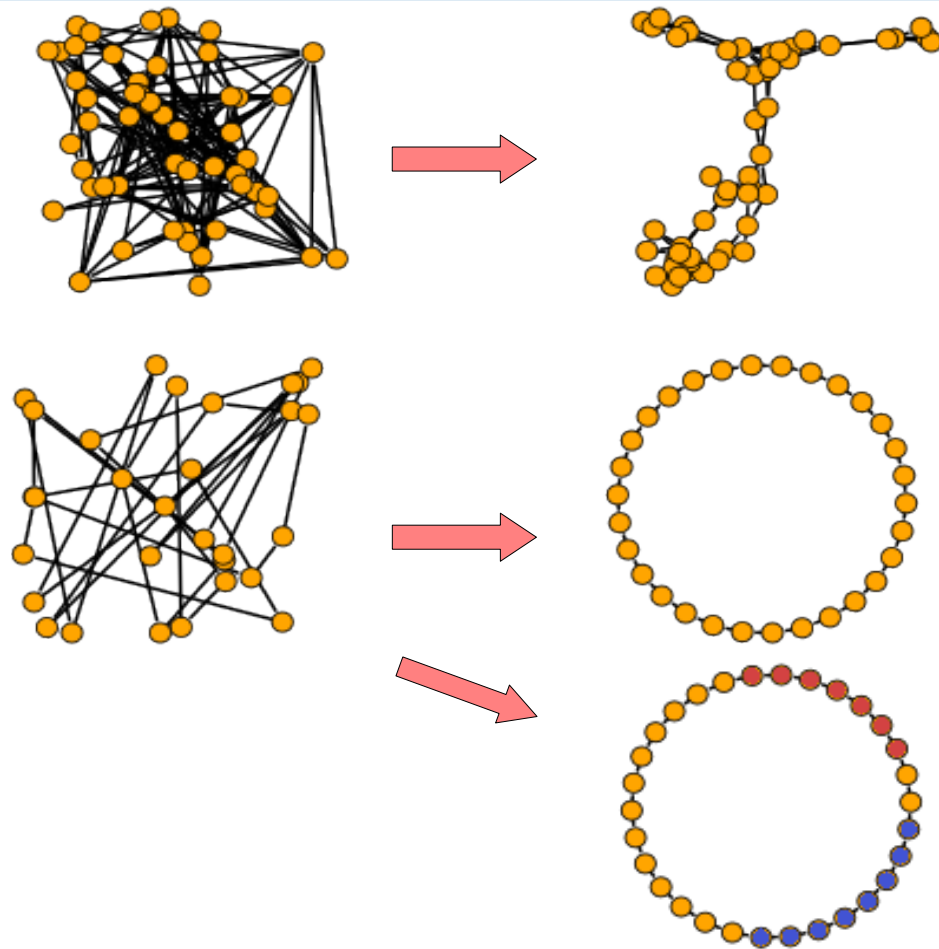


A **community**, with respect to graphs, can be defined as a subset of nodes that are densely connected to each other and loosely connected to the nodes in the other communities in the same graph

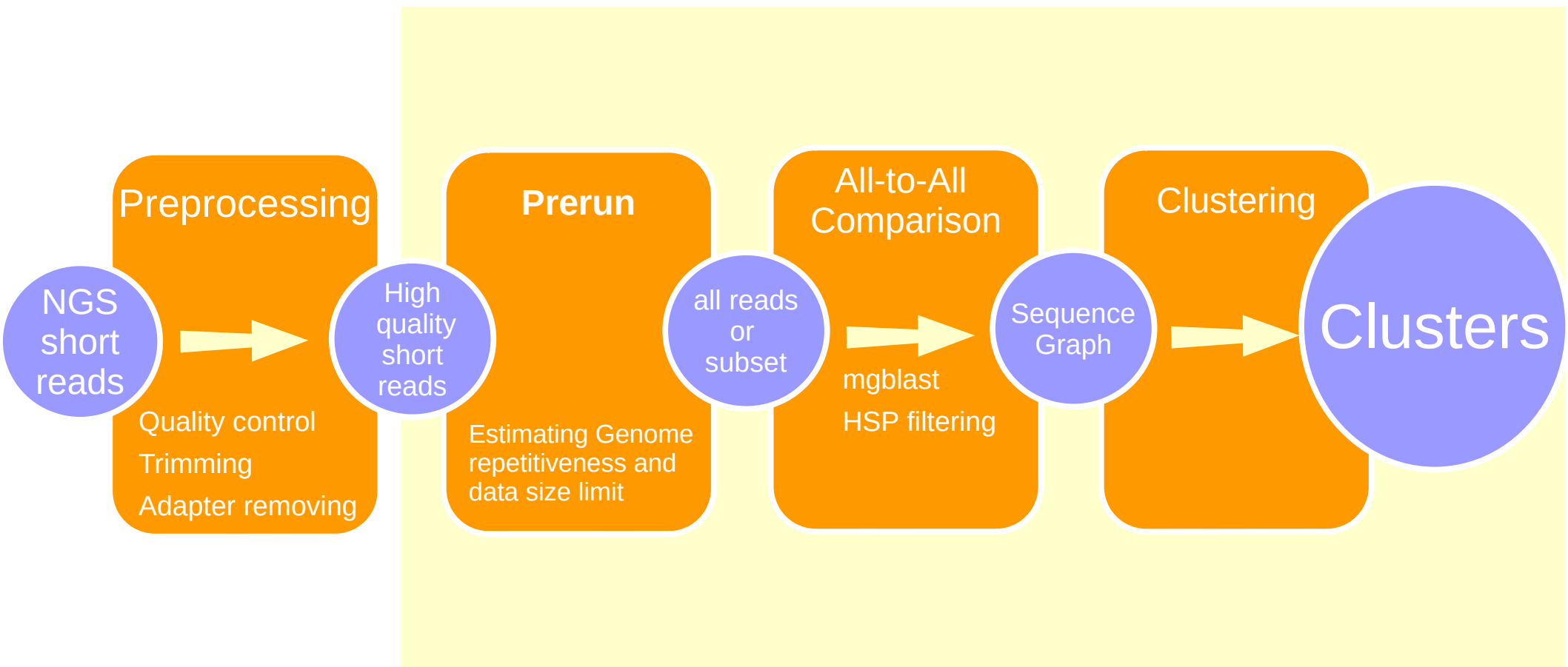
community ~ cluster ~ repeat family

Graph Based Representation of Sequence Reads

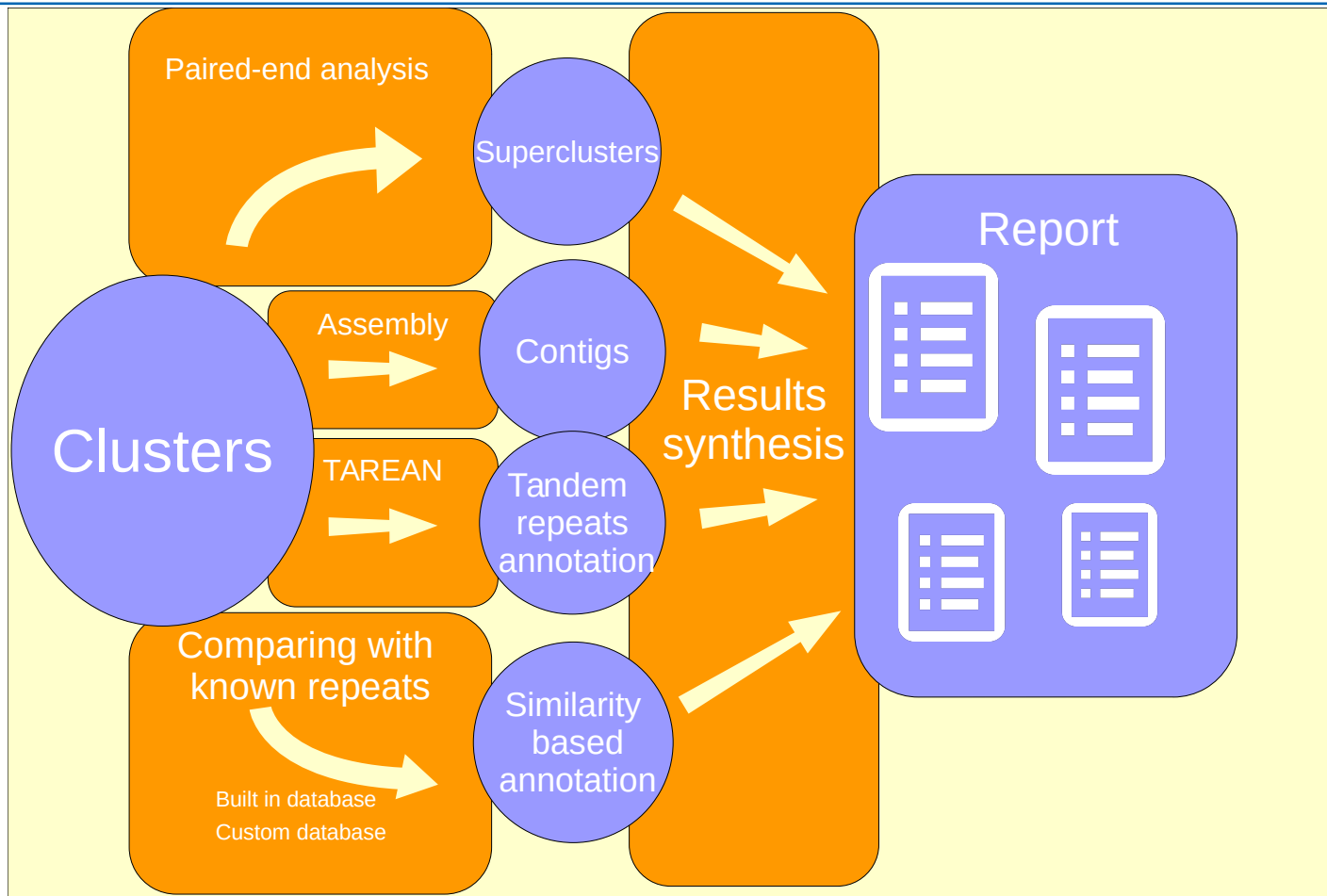
- Informative graphical representation
- Graph layout
- Vertex coloring



RepeatExplorer Pipeline



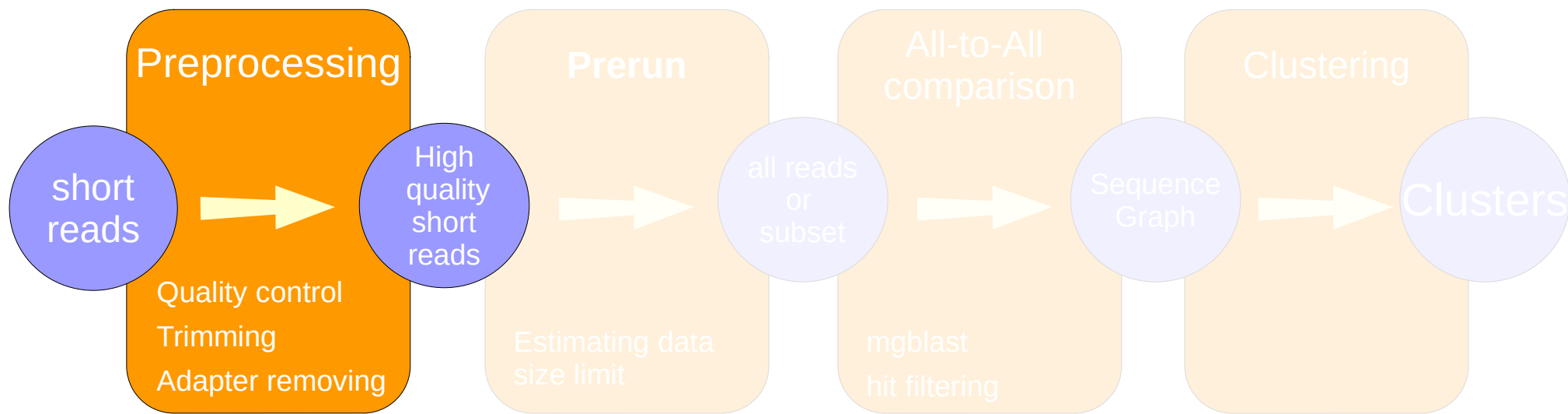
RepeatExplorer Pipeline



RepeatExplorer pipeline

Input data

- Short reads – hundreds of nt
- Paired-end, interleaved
- Single-end
- Pre-processed
- Uniform length
- FASTA format

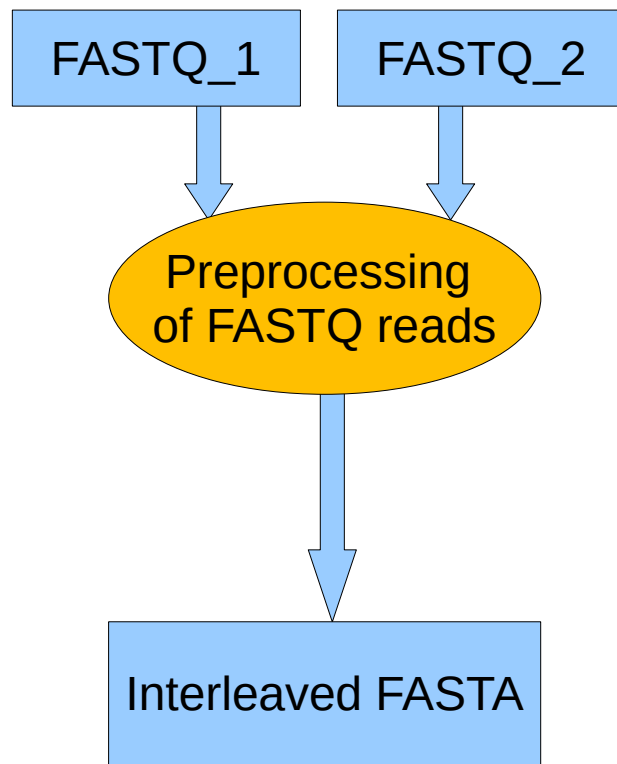


Preprocessing

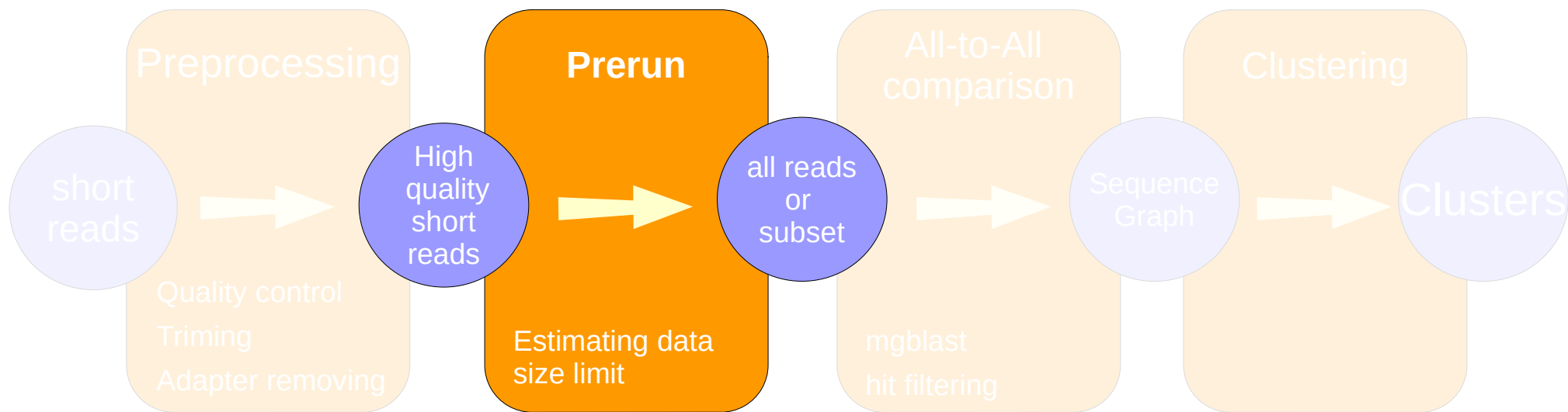
RepeatExplorer utilities:

Preprocessing of FASTQ reads

1. Trimming (optional)
2. Filter by quality
3. Cutadapt filtering
4. Discard single reads, keep complete pairs
5. Sampling (optional)
6. Interlacing

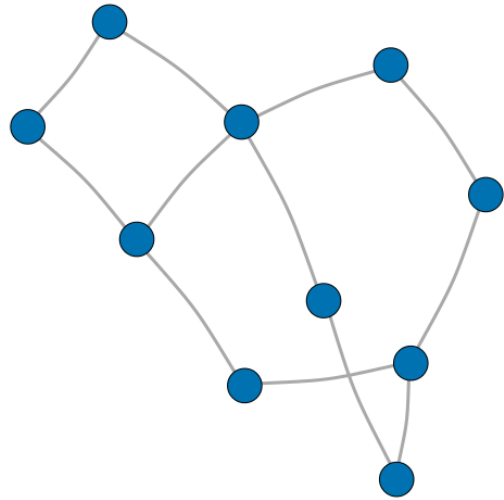


RepeatExplorer pipeline

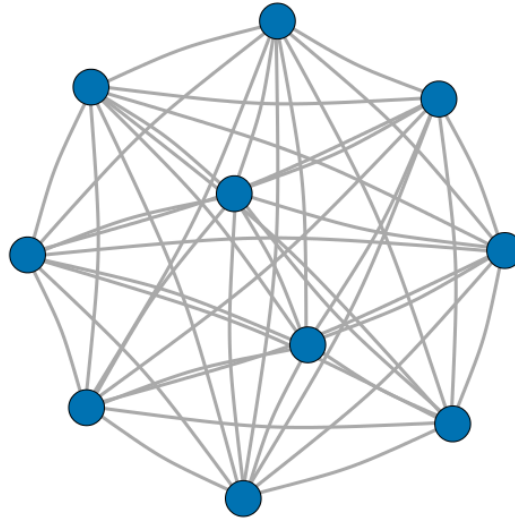


Prerun

All-to-all sequence comparison on small sample of input data



$$\begin{aligned} N &= 10 \\ E &= 12 \\ k_g &= 0.27 \end{aligned}$$



$$\begin{aligned} N &= 10 \\ E &= 45 \\ k_g &= 1.0 \end{aligned}$$

graph density - k_g is genome specific coefficient and depends on the repetitive content and genome size

Density corresponds to probability that two randomly taken sequences from genome will be similar

k_g is used to estimate maximum number of processable reads

$$k_g = \frac{2E}{N(N-1)}$$

Prerun

All-to-all sequence comparison on small sample of input data

All-to-all sequence comparison on small sample of NGS reads

$$k_g = \frac{2E}{N(N-1)}$$

N .. 20,000 sample reads

E .. number of identified similarity hits

k_g is used to estimate maximum number of reads **N_{max}** providing that we can process with available RAM (**M**)

$$N_{max} = \sqrt{m \frac{M}{k_g}}$$

Prerun

All-to-all sequence comparison on small sample of input data

All-to-all sequence comparison on small sample of NGS reads

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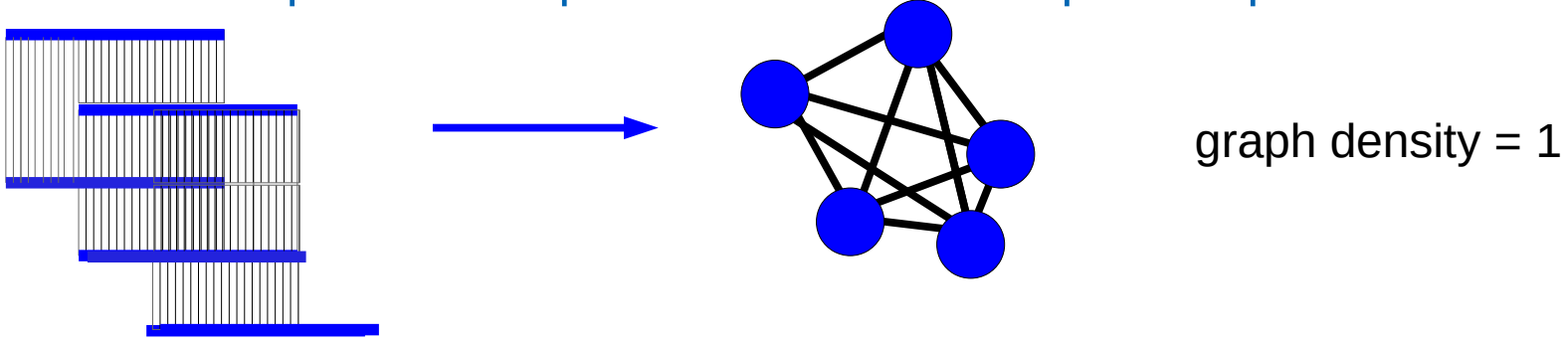
$$N_{max} = \sqrt{m \frac{M}{k_g}}$$

Species	Number of reads	Genome Size (1C)	Coverage [%]
<i>Musa acuminata</i>	3,046,164	623 Mbp	48.9
<i>Lasiurus borealis</i>	4,256,140	2,526 Mbp	16.8
<i>Pisum sativum</i>	3,011,839	4,300 Mbp	7.0
<i>Vicia panonica</i>	1,039,442	5,730 Mbp	1.8
<i>Silene latifolia</i>	2,943,062	5,850 Mbp	5.0
<i>Secale cereale</i>	1,899,753	7,917 Mbp	2.4
<i>Lathyrus latifolius</i>	1,464,940	9,980 Mbp	1.5
<i>Fritilaria imperialis</i>	12,220,382	42,400 Mbp	2.9
<i>Fritilaria affinis</i>	1,168,248	45,000 Mbp	0.3

Number of reads which can be processed with 16GB RAM

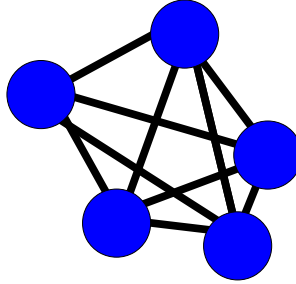
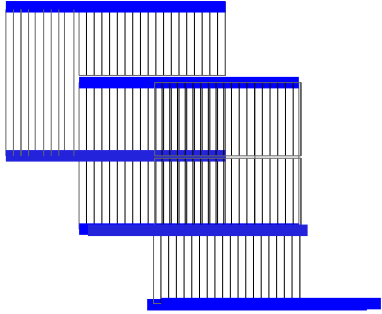
Prerun

- All-to-all sequence comparison on small sample of input data



Prerun

All-to-all sequence comparison on small sample of input data



graph density = 1



graph density = 0.54

Prerun

All-to-all sequence comparison on small sample of input data



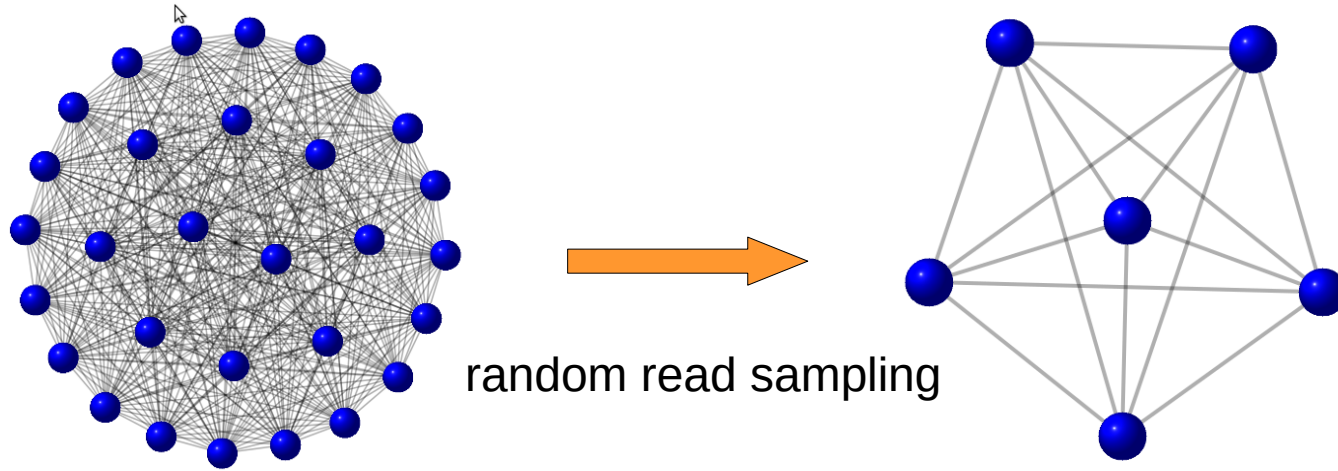
Number of reads (Vertices)	44,772
Number of similarity hits (Edges)	542,348,907

Input data (All reads)	2,000,000
Total number of similarity hits	1,394,970,205

Approx 1/3 of stored similarity hits originate from satellite which represent only 2% of genome

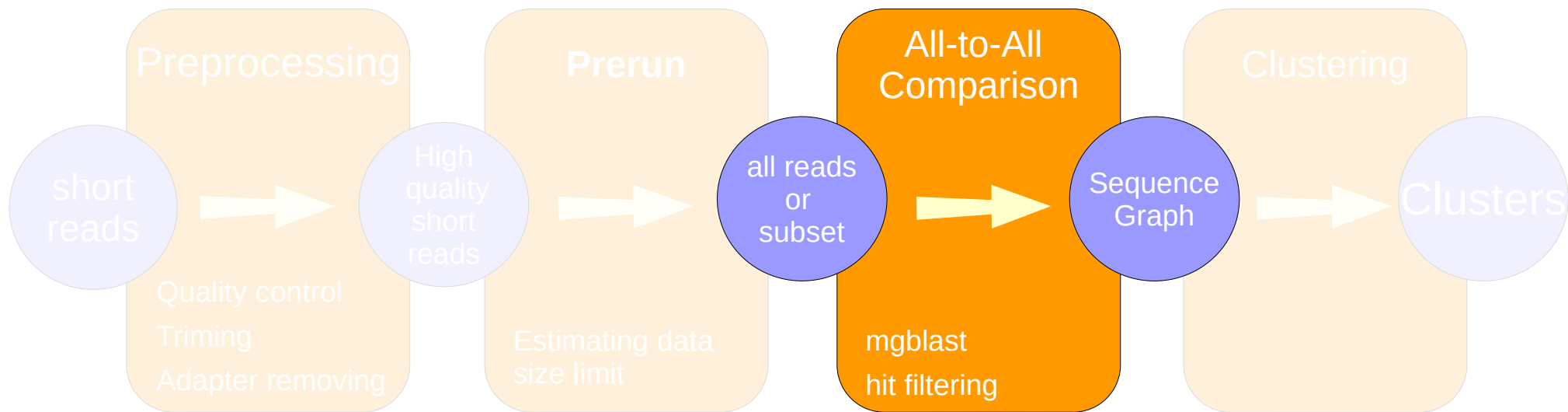
Prerun

Satellite filtering (optional)



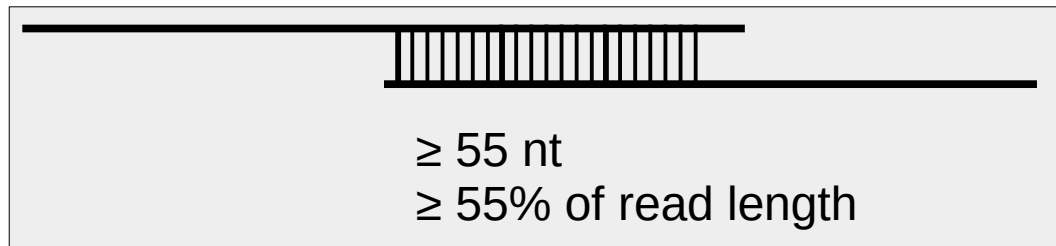
- Clusters composed from satellite reads can be scaled down without losing information.
- Sample of 10% of reads of is kept in analysis to keep track of this satellite

RepeatExplorer pipeline



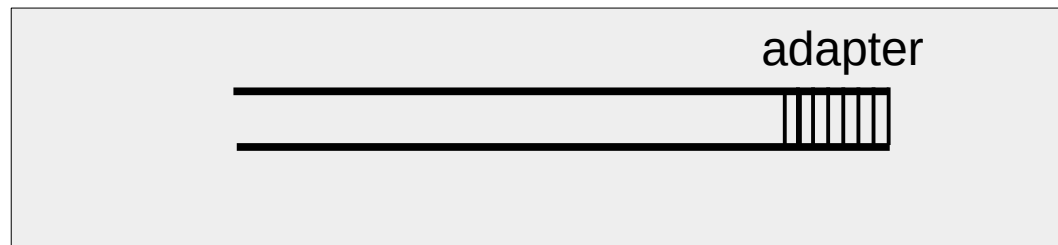
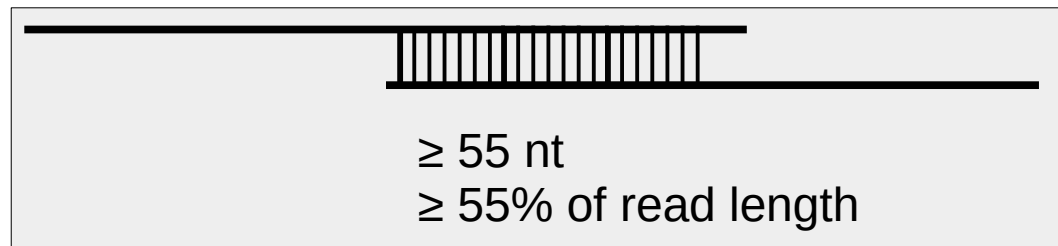
All-to-all comparison

- Similarity search using **mgblast**
- Default threshold:
 - overlap : 55 nt and 55% of the length
 - minimal similarity 90%
- By default mgblast is using **DustMasker** (low complexity repeat filter)
 - simple repeats are underestimated or not detected (e.g. telomeric motifs, microsatellites)
 - Masking of low complexity can be disabled → long running time and increased memory usage



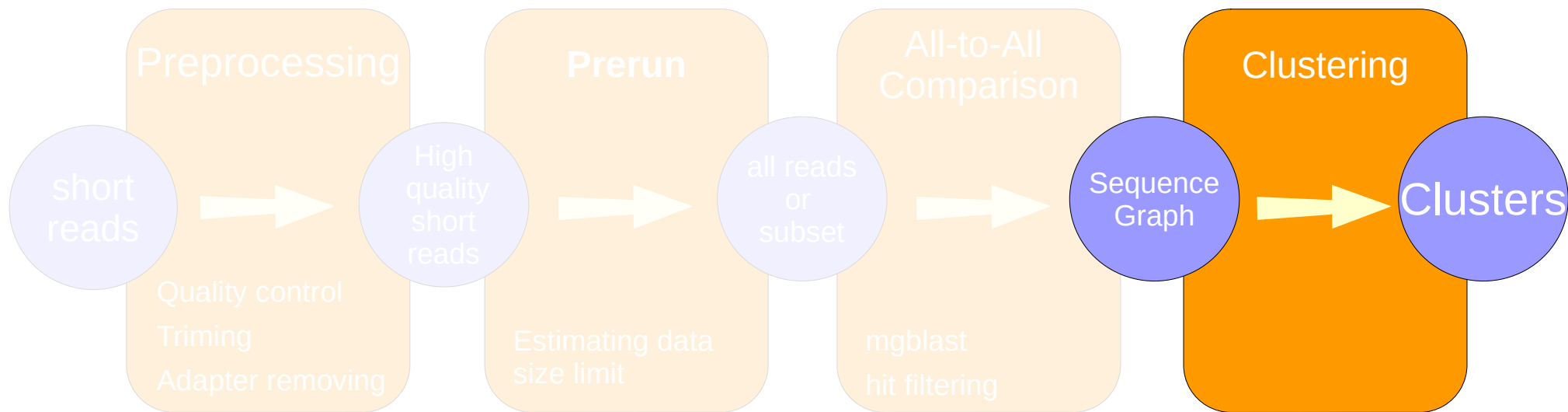
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- Adapters in sequence can slow down all-to-all search

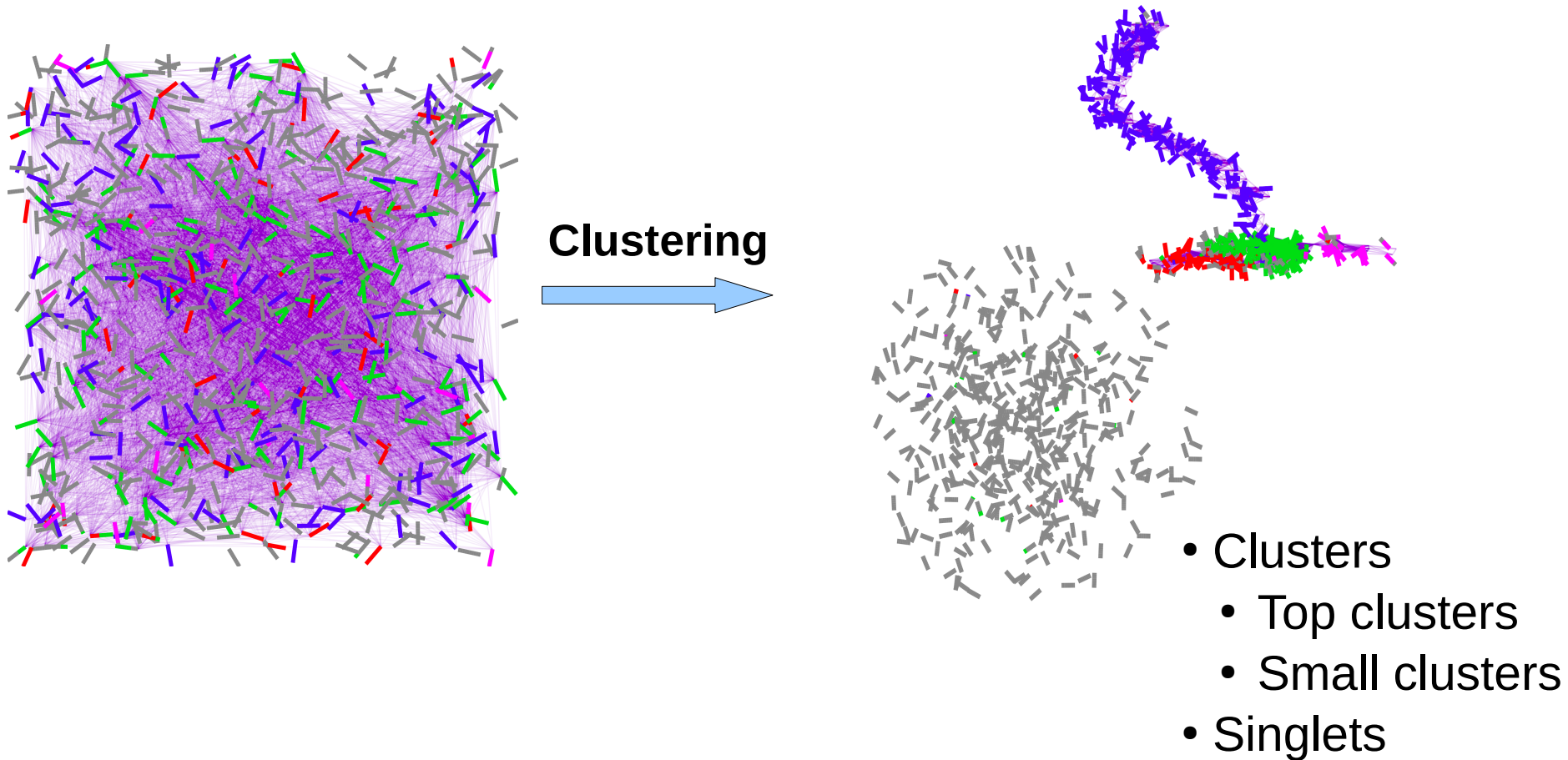


RepeatExplorer pipeline

- Graph is divided into subgraphs (clusters/communities)
- Clusters have dense connections between the nodes within the clusters but sparse connections between nodes in different clusters

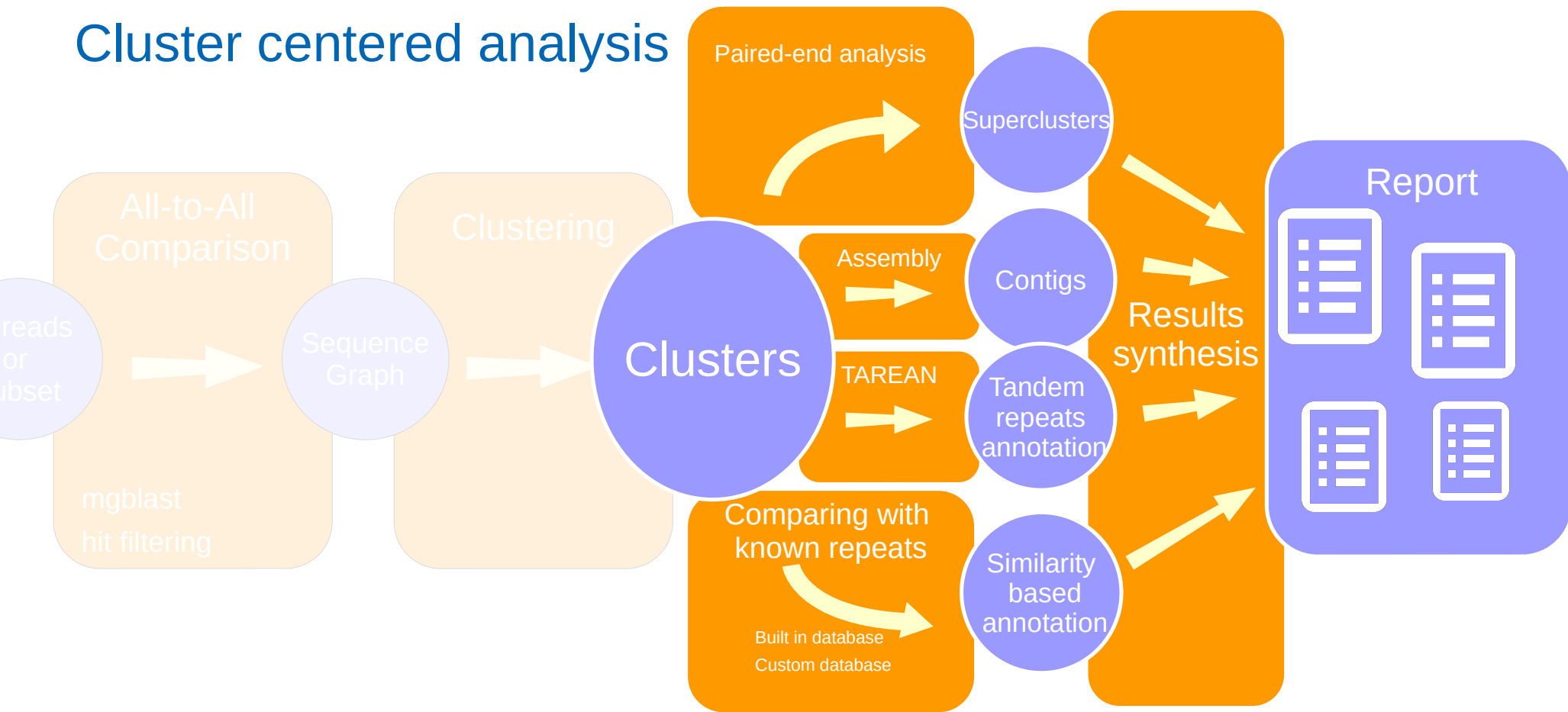


RepeatExplorer pipeline

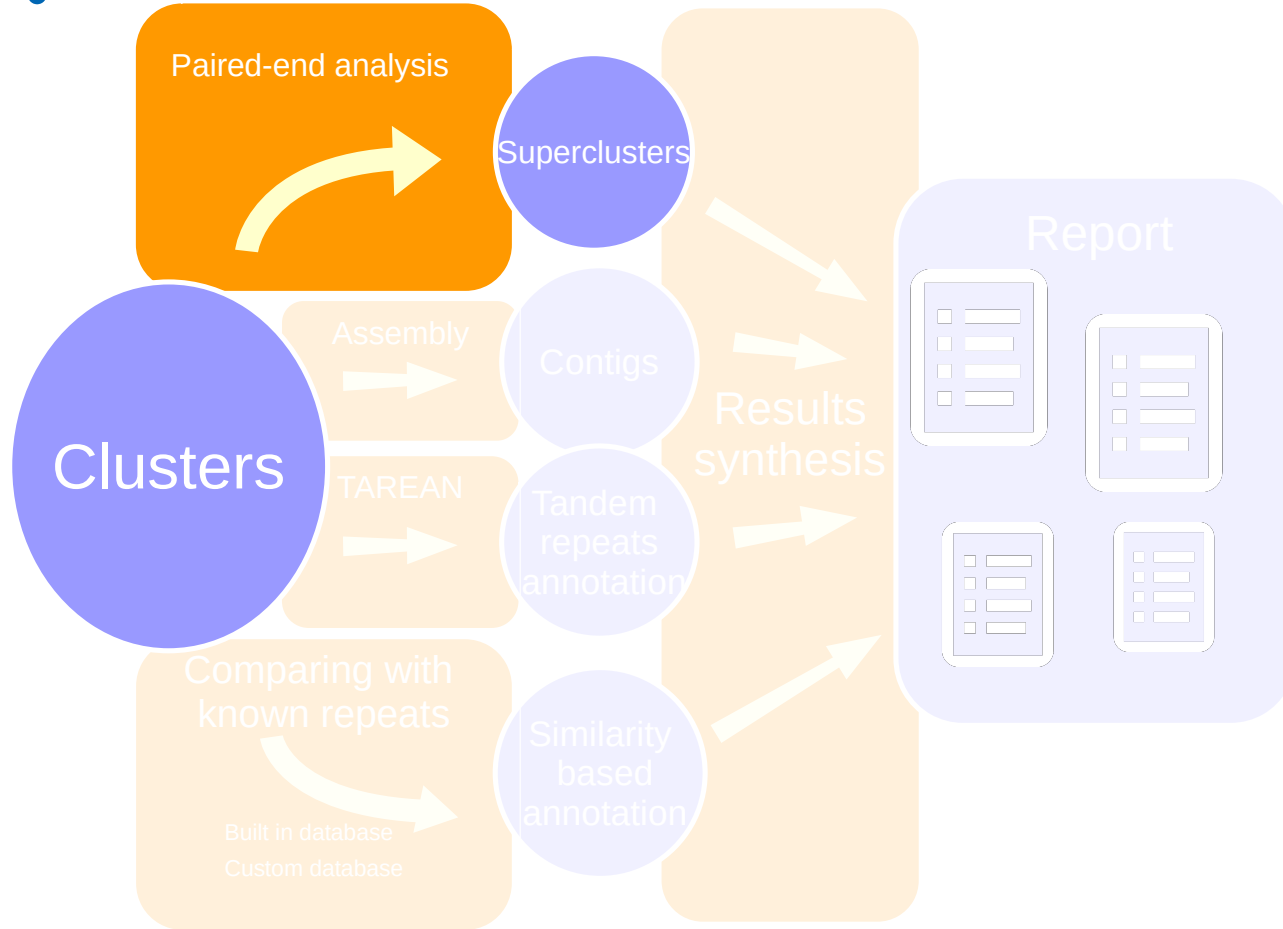


RepeatExplorer pipeline

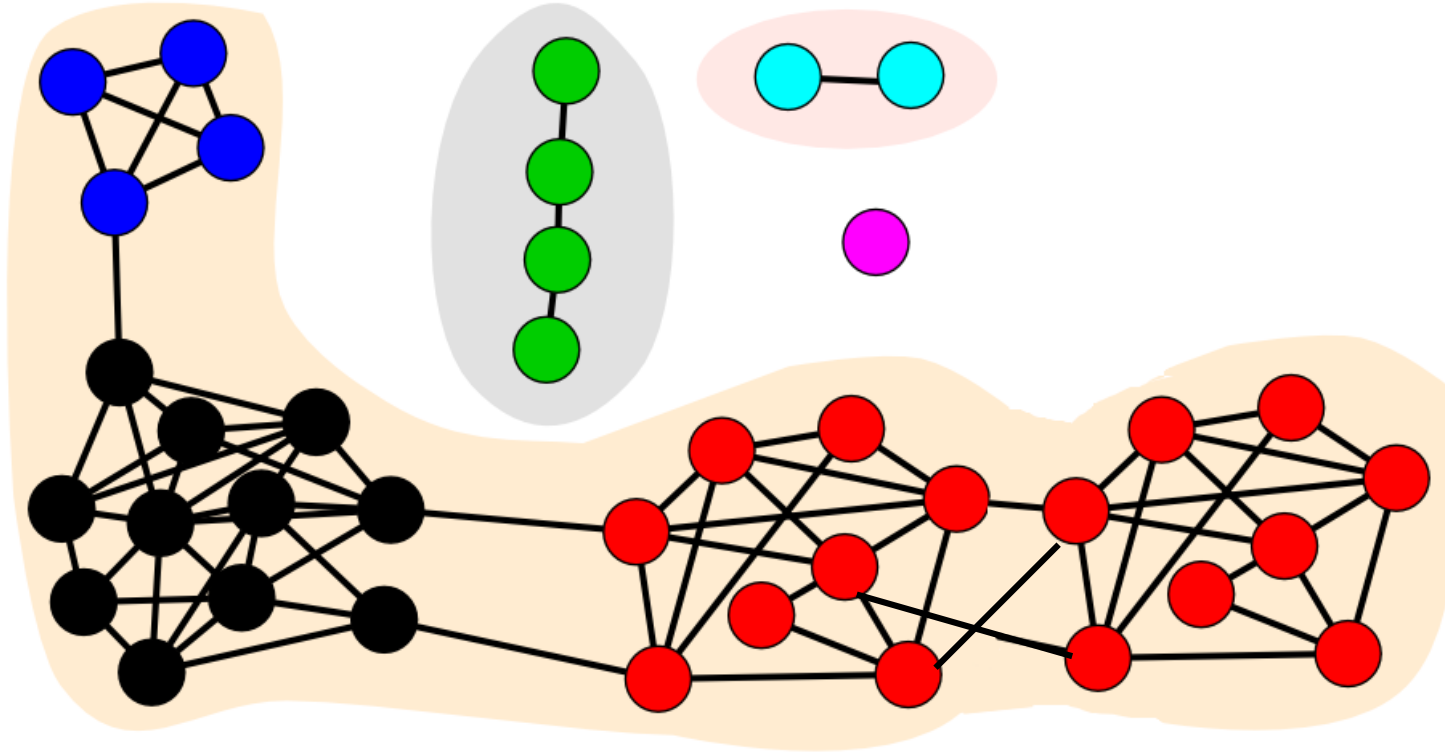
Cluster centered analysis



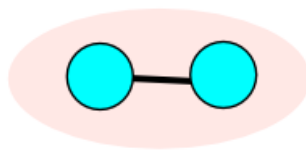
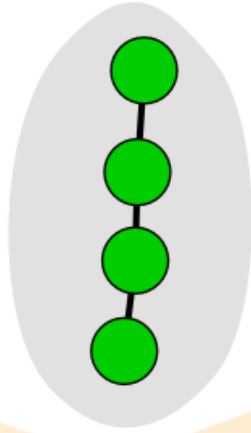
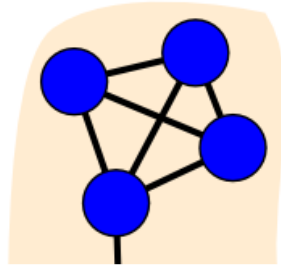
RepeatExplorer pipeline



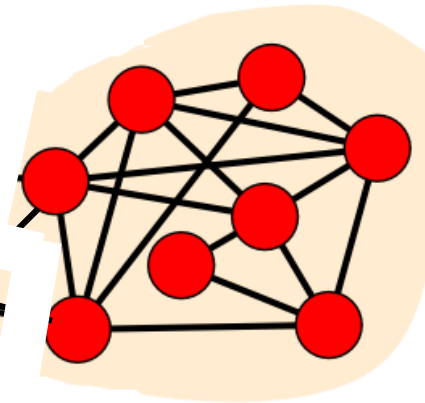
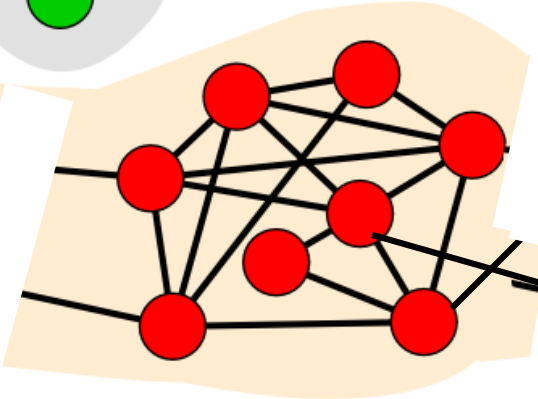
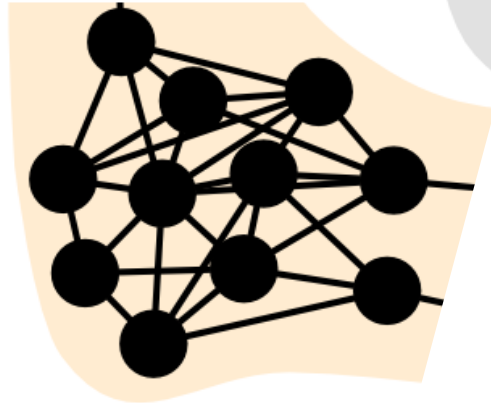
Clusters and Superclusters



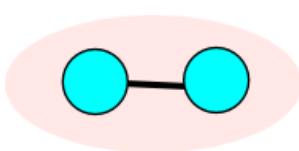
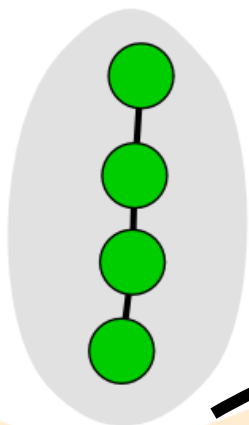
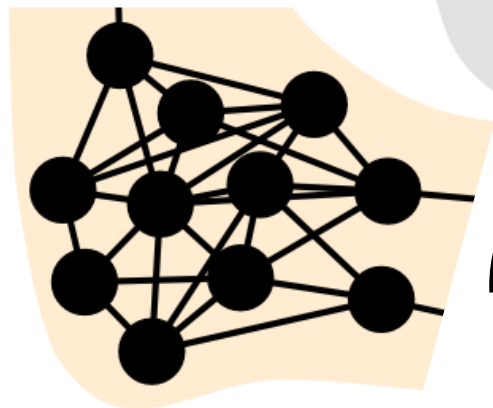
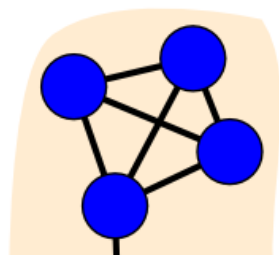
Clusters and Superclusters



Sometimes (often) reads which belong to single repeat family are split into multiple clusters

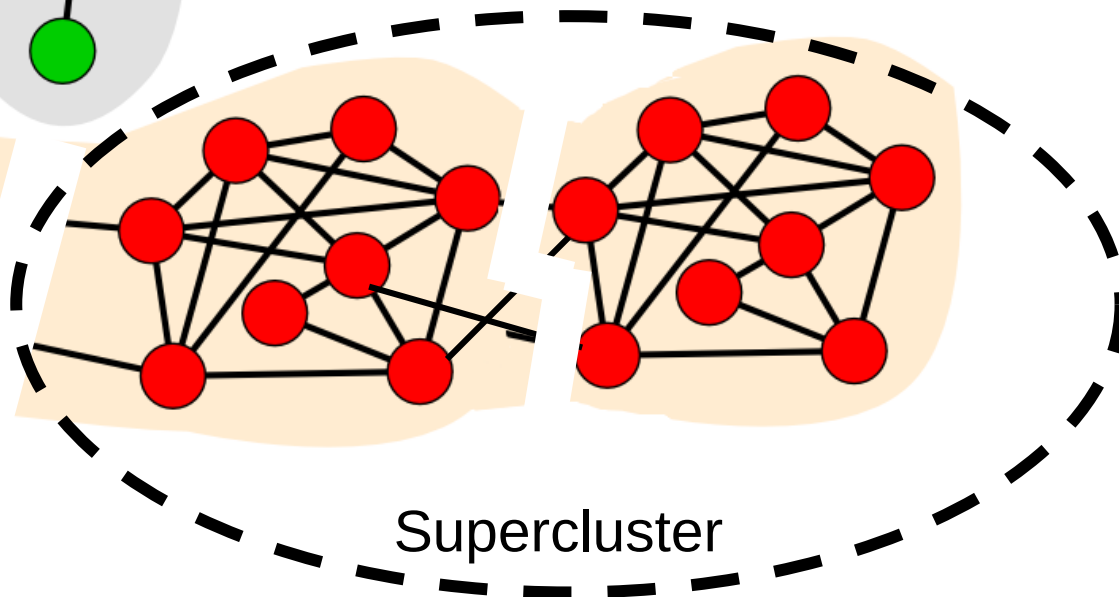


Clusters and Superclusters



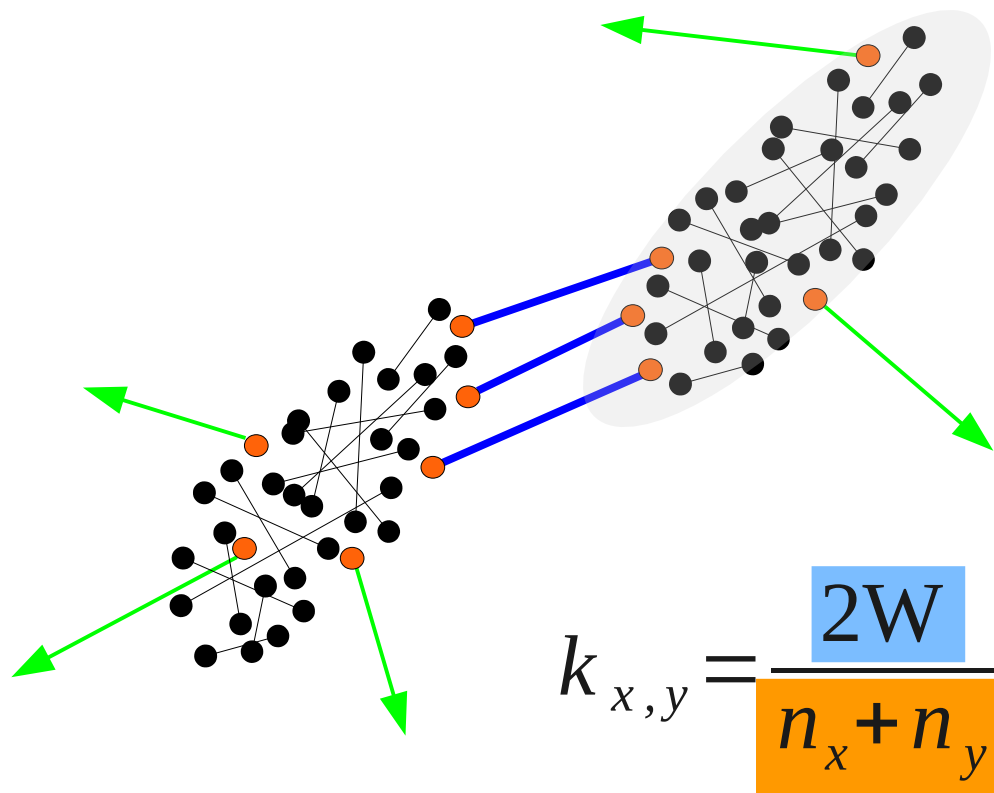
Sometimes (often) reads which belong to single repeat family are split into multiple clusters

We need to identify such false splits



Clusters and Superclusters

Identification of supercluster using paired-end reads



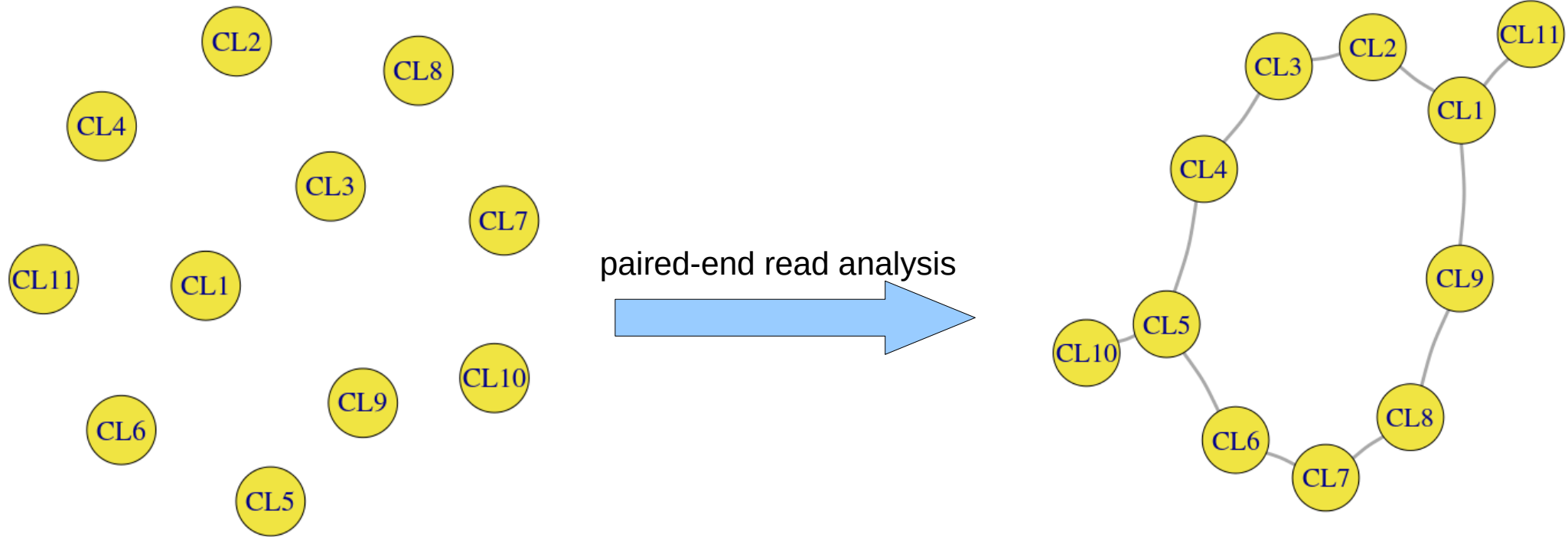
W number of reads pairs shared between clusters x and y
 n_x and n_y is number of reads in cluster x and cluster y with absent read mate within the same cluster respectively

Suitable $k_{x,y}$ cutoff 0.05 – 0.2
full connection: $k_{x,y}=1$
no connection $k_{x,y}=0$

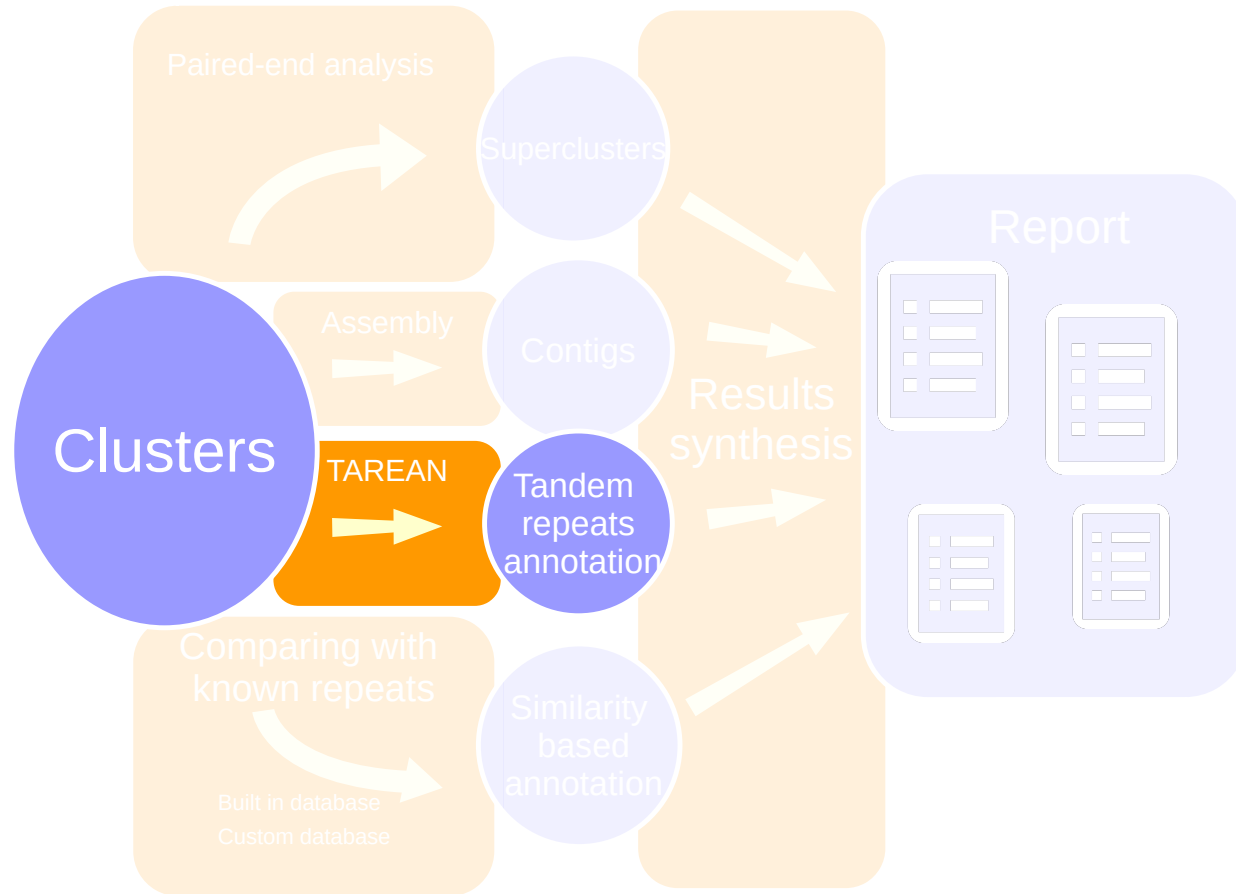
$$k_{x,y} = \frac{2W}{n_x + n_y}$$

Clusters and Superclusters

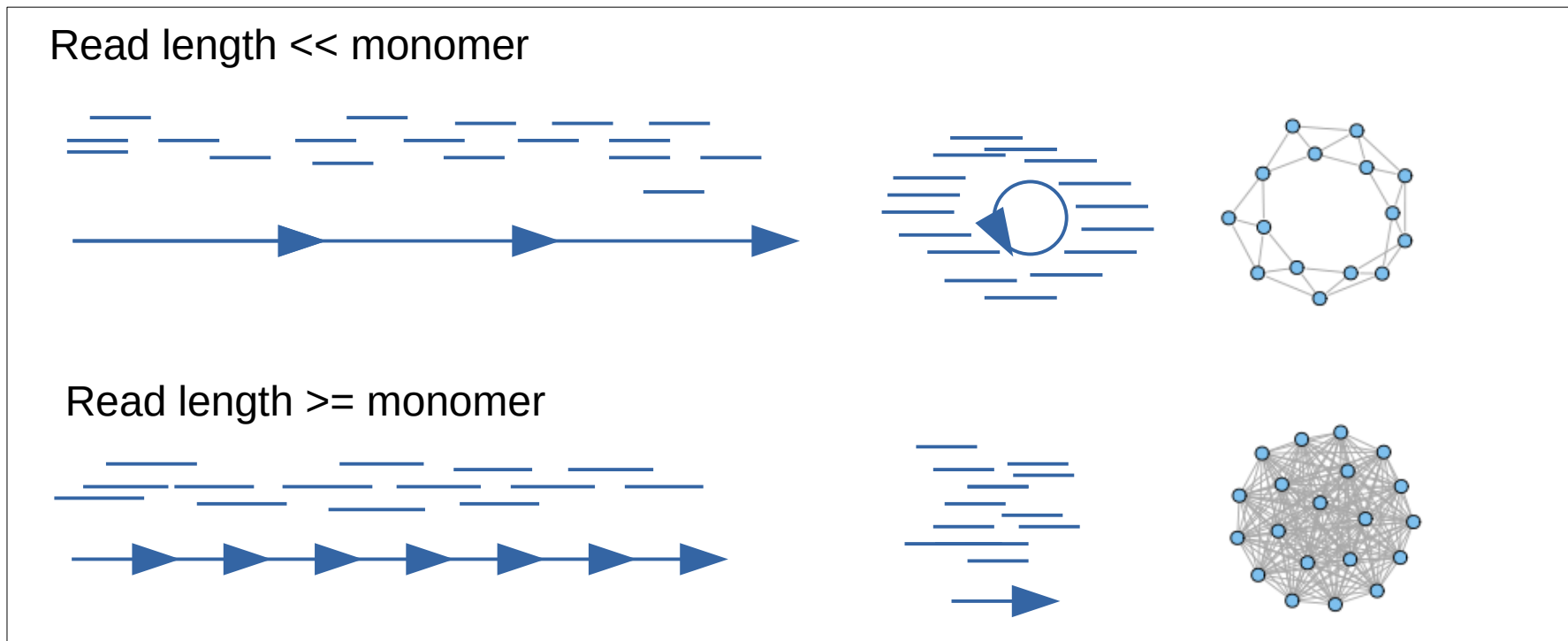
Identification of supercluster using paired-end reads



RepeatExplorer pipeline



Tandem Repeat Analyzer - TAREAN

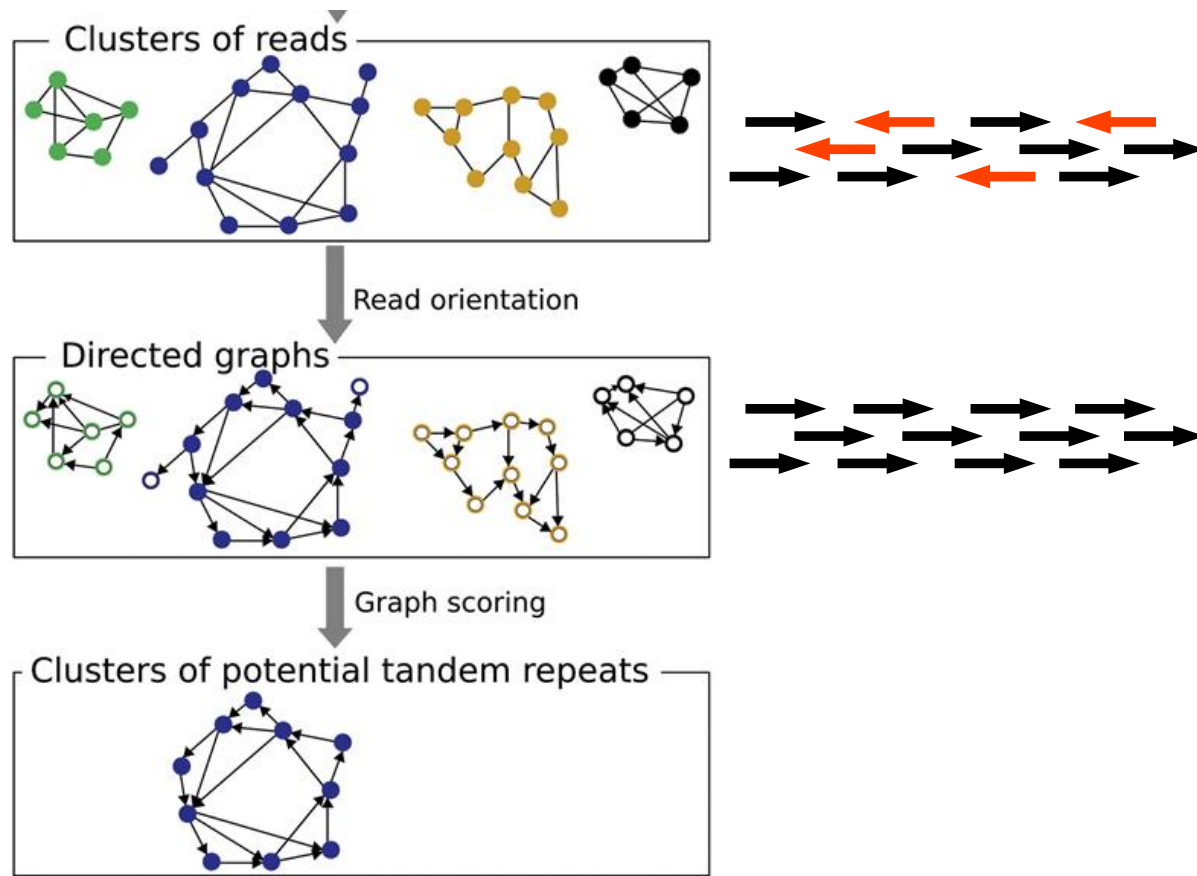


TAREAN calculates **graph layout** and provide automatic analysis of **graph topology** with the aim to identify **tandem repeats**

Tandem Repeat Analyzer - TAREAN

Principle

- All reads within cluster are set to have same 3'to5' orientation with hypothetical tandem repeat monomer
- Directed graph is constructed
- Large circular structures are detected (a.k.a strongly connected component)
- **Connected Component index**

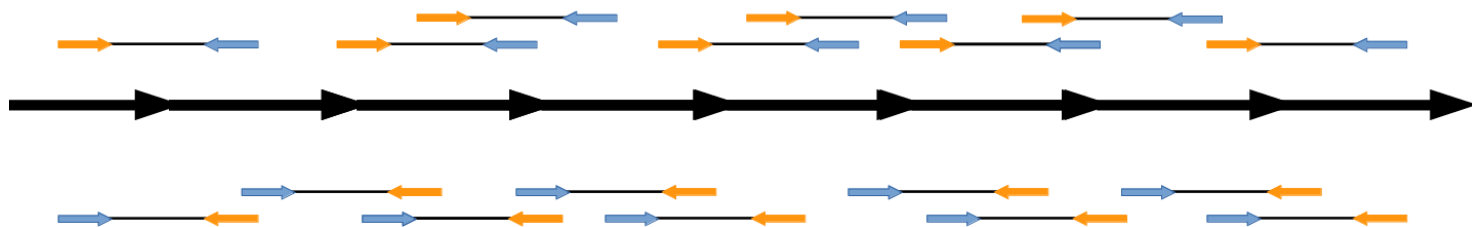


Tandem Repeat Analyzer - TAREAN

Principle

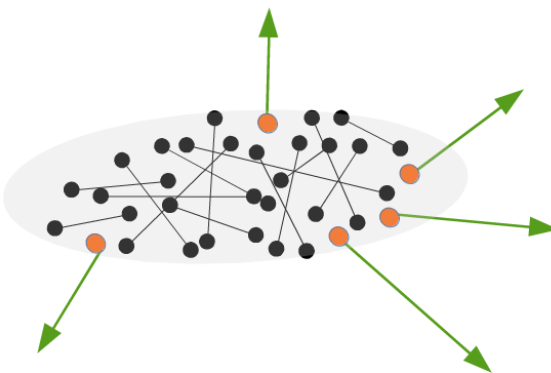
Paired-End Sequencing

Forward
Reverse



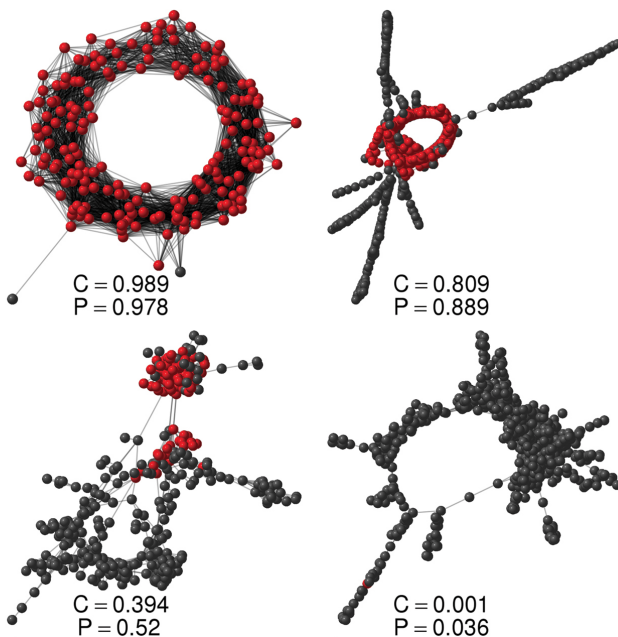
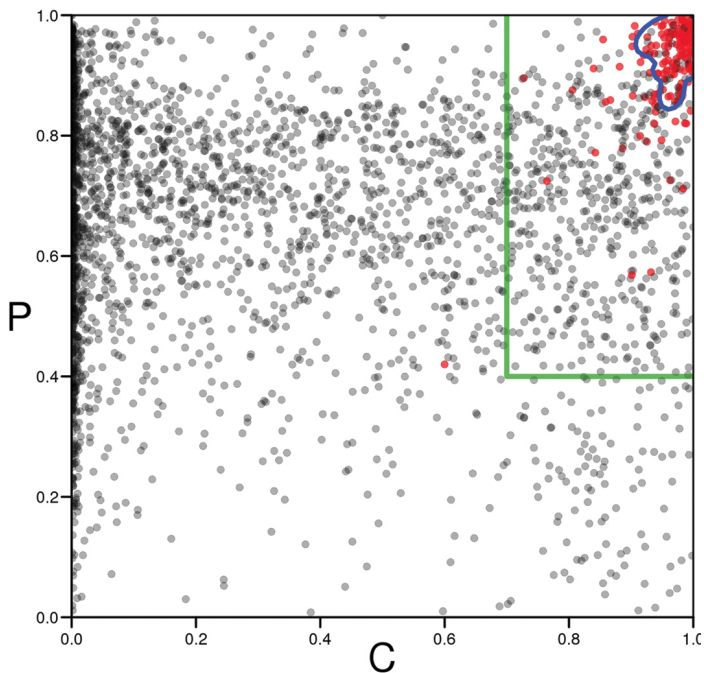
Pair completeness = fraction of complete pairs in cluster

In clusters which are derived from tandem repeat, most of the paired-end reads should be complete



Tandem Repeat Analyzer - TAREAN

Principle



Five groups of clusters by TAREAN

- **Putative satellite (high confidence)**
high P and C score
- **Putative satellite (low confidence)**
P and C score lower
- **Putative LTR element**
Primer binding site detected, presence of long ORF
- **rDNA**
tandem organization + similarity to known rDNA sequences
- **Other clusters**
no tandem repeat like structure

RepeatExplorer pipeline

Principle

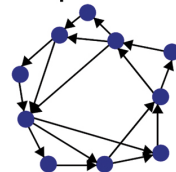
Reconstruction of tandem repeat monomer

- k-mer based approach
- multiple variants reported
- sorted based on significance

TAREAN limitation

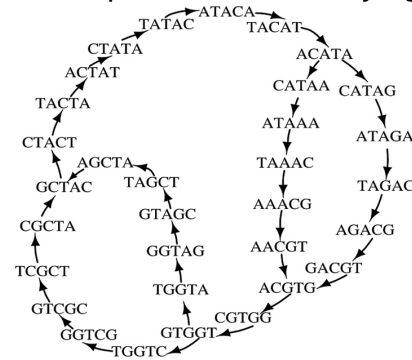
- paired end reads required
- limited sensitivity to TR with very short monomer

Clusters of potential tandem repeats



k-mer counting

Tandem repeats as de Bruijn graphs

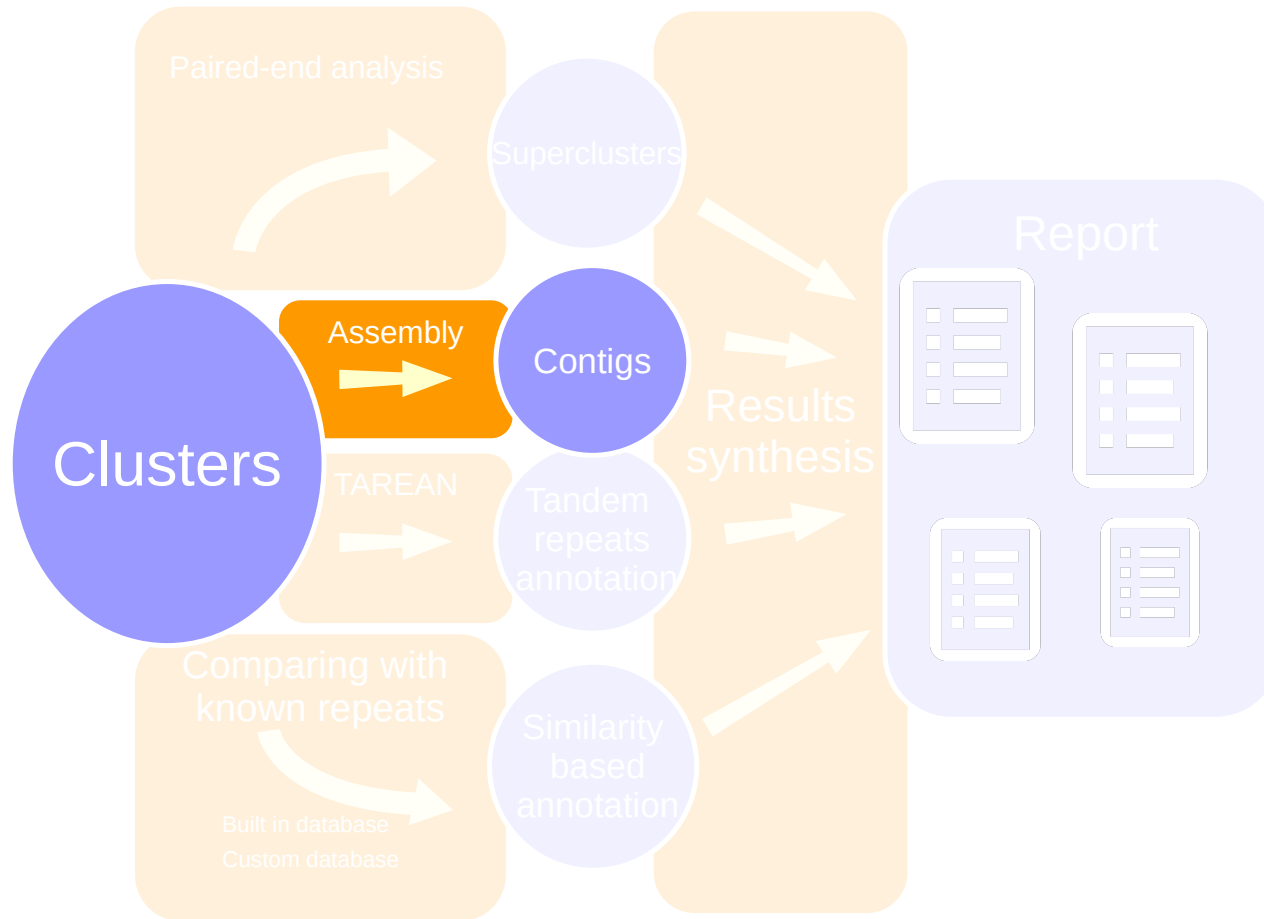


Identification of cycles

Consensus sequences

CATAGACGTGGTGGCTACTATA

RepeatExplorer pipeline



Contig assembly

Reads are assembled by CAP3 program, each cluster separately:

ACTGTGTCGTCGTCGTCGTGTG

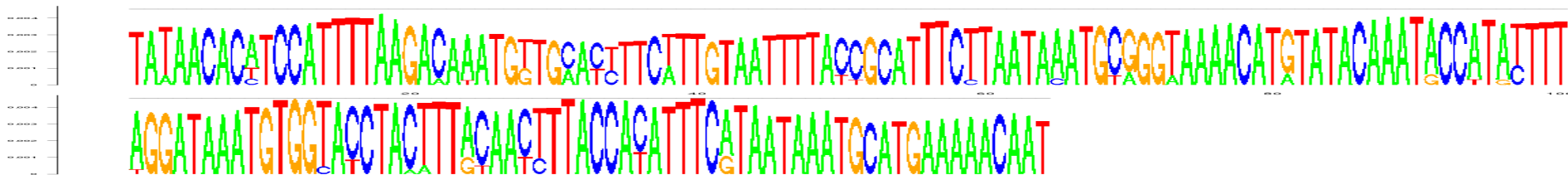
CGTCGTCG-CGTGTGGT

GTCGTGTG-TTGTCGTCTGA

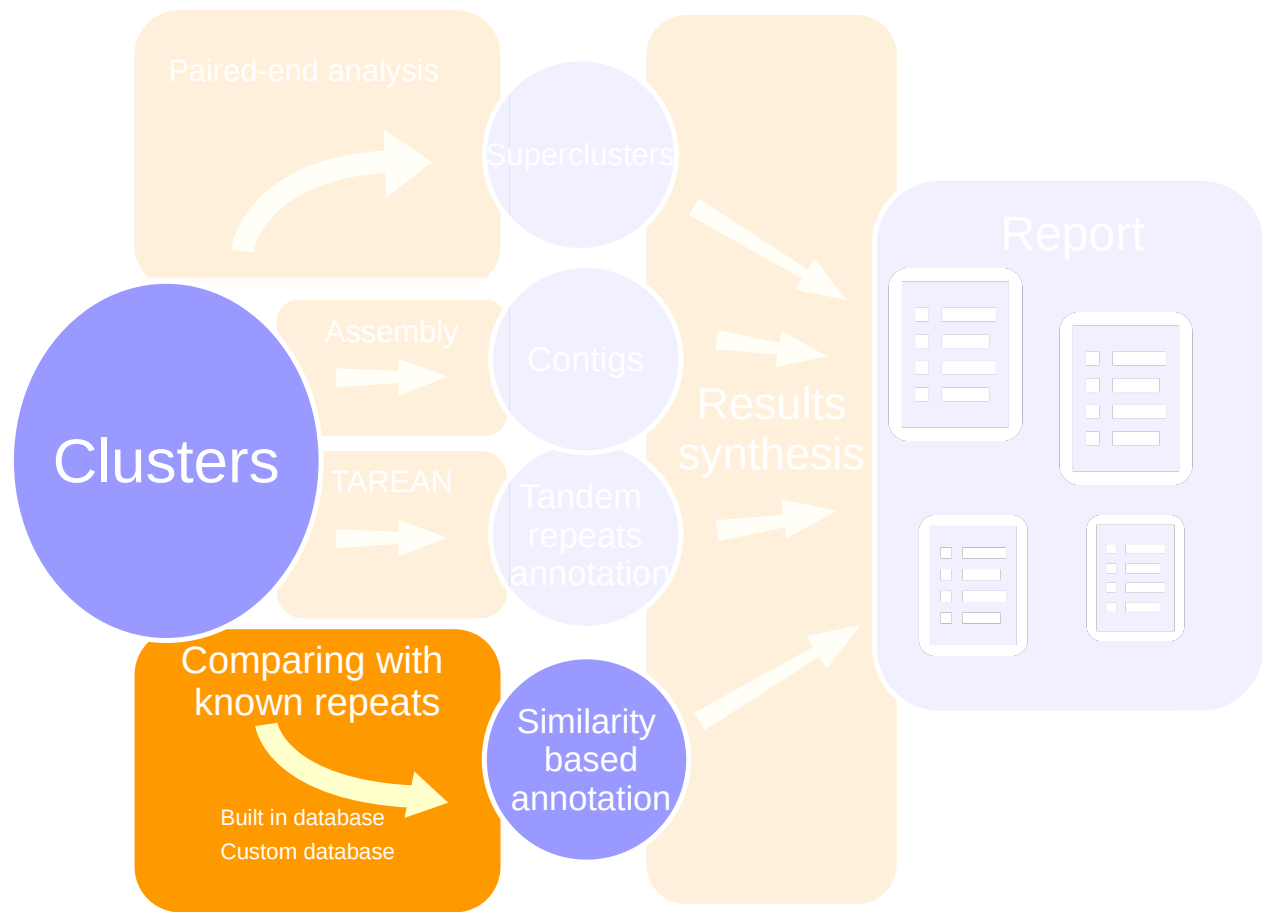
Reads

ACTGTGTCGTCGTCGTCGTGTGGTTGTCGTCTGA **Contig**

High confidence putative satellite clusters are not assembled by CAP3, instead TAREAN generate **k-mer based** consensus:



RepeatExplorer pipeline



All **reads** are compared with:

- Database of protein domains (REXdb)
- DNA database
 - rDNA, tRNA
 - Organelle DNA
 - potential contaminants
- Custom database (optional)

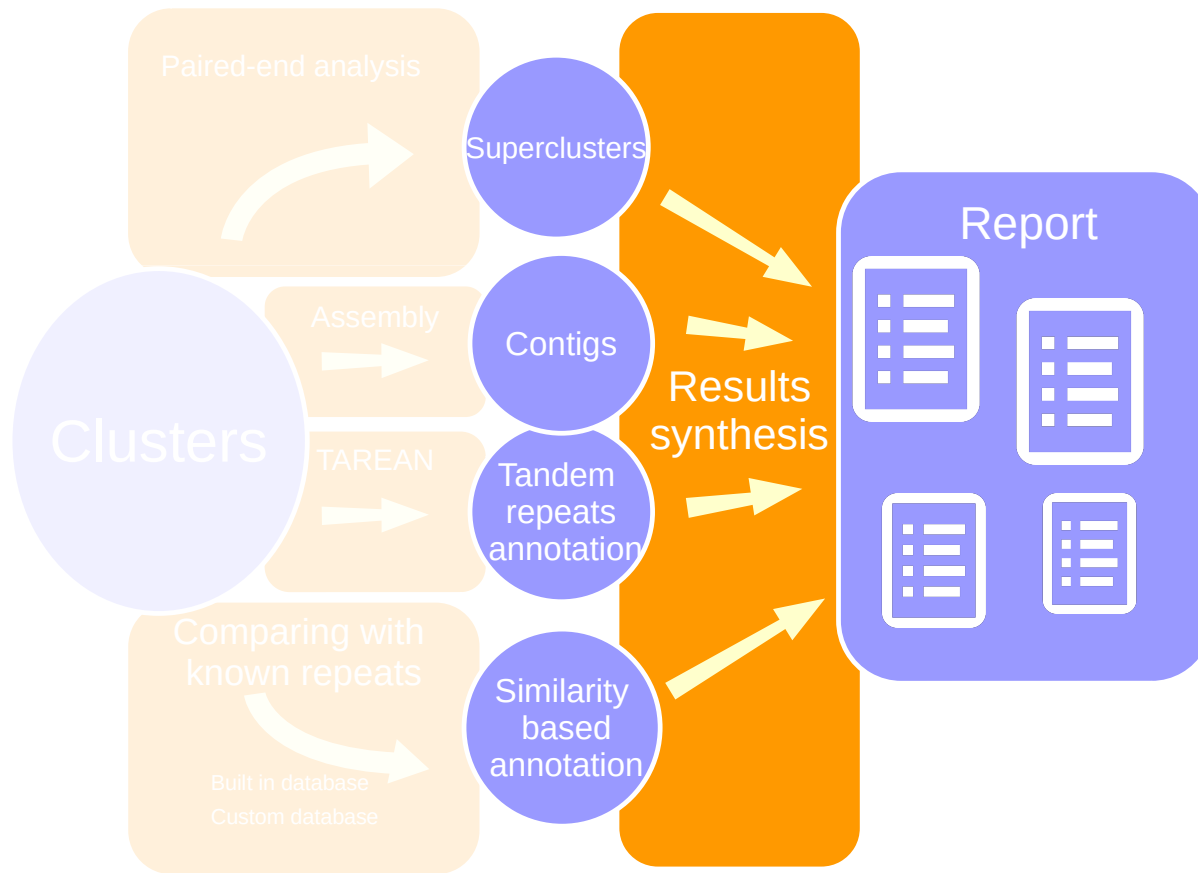
Similarity search

Database sequence classification	Protein domain	Number of reads with similarity hit	Proportion No of reads / cluster size
mitochondria		25	0.0023
Ogre Ty3-RH	Ty3-RH	2977	0.27402
Retand Ty3-RH	Ty3-RH	2	0.00018
Ogre Ty3-RT	Ty3-RT	3473	0.31968
Ogre Ty3-aRH	Ty3-aRH	1713	0.15768

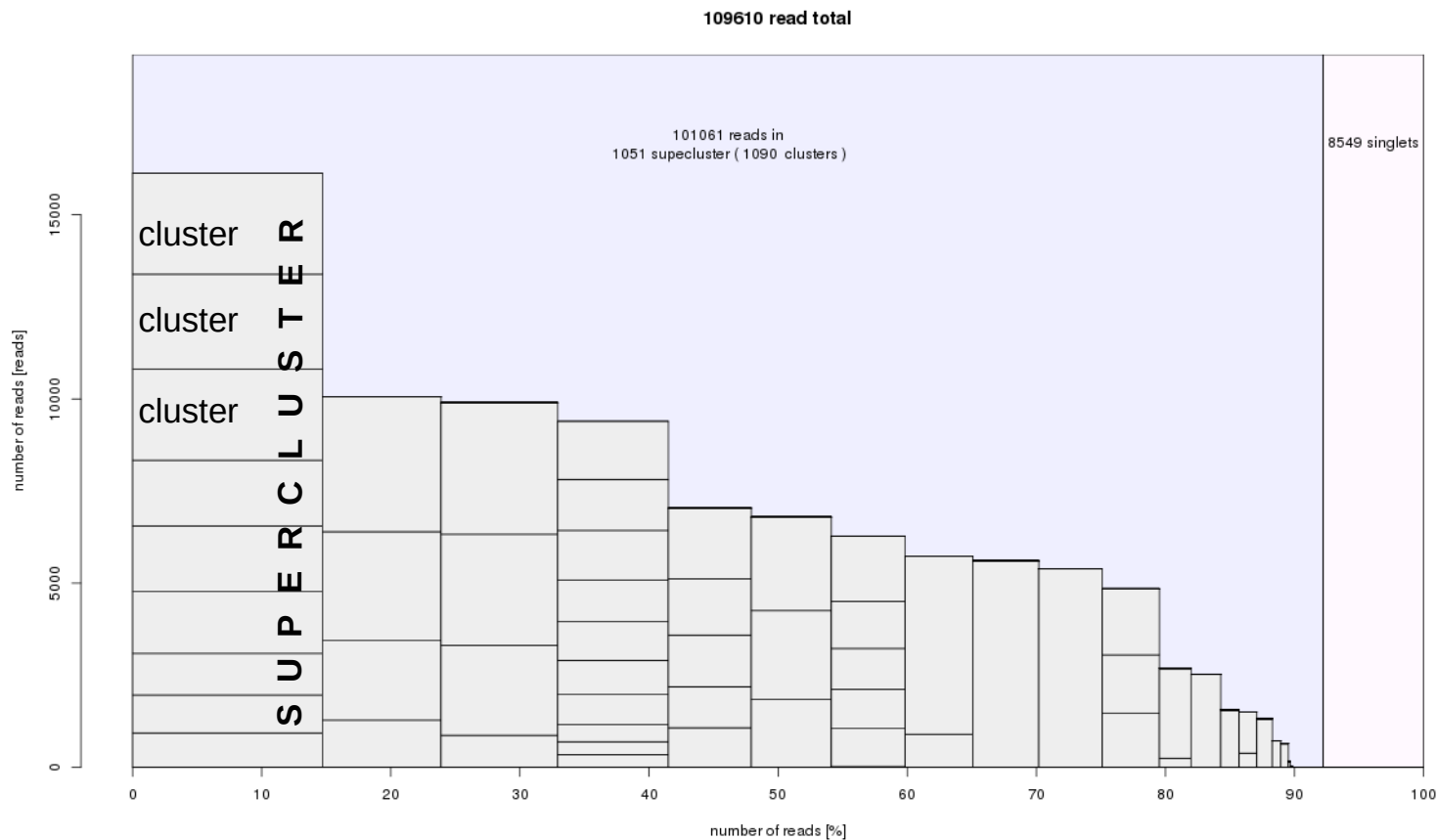
RepeatExplorer pipeline

Reporting:

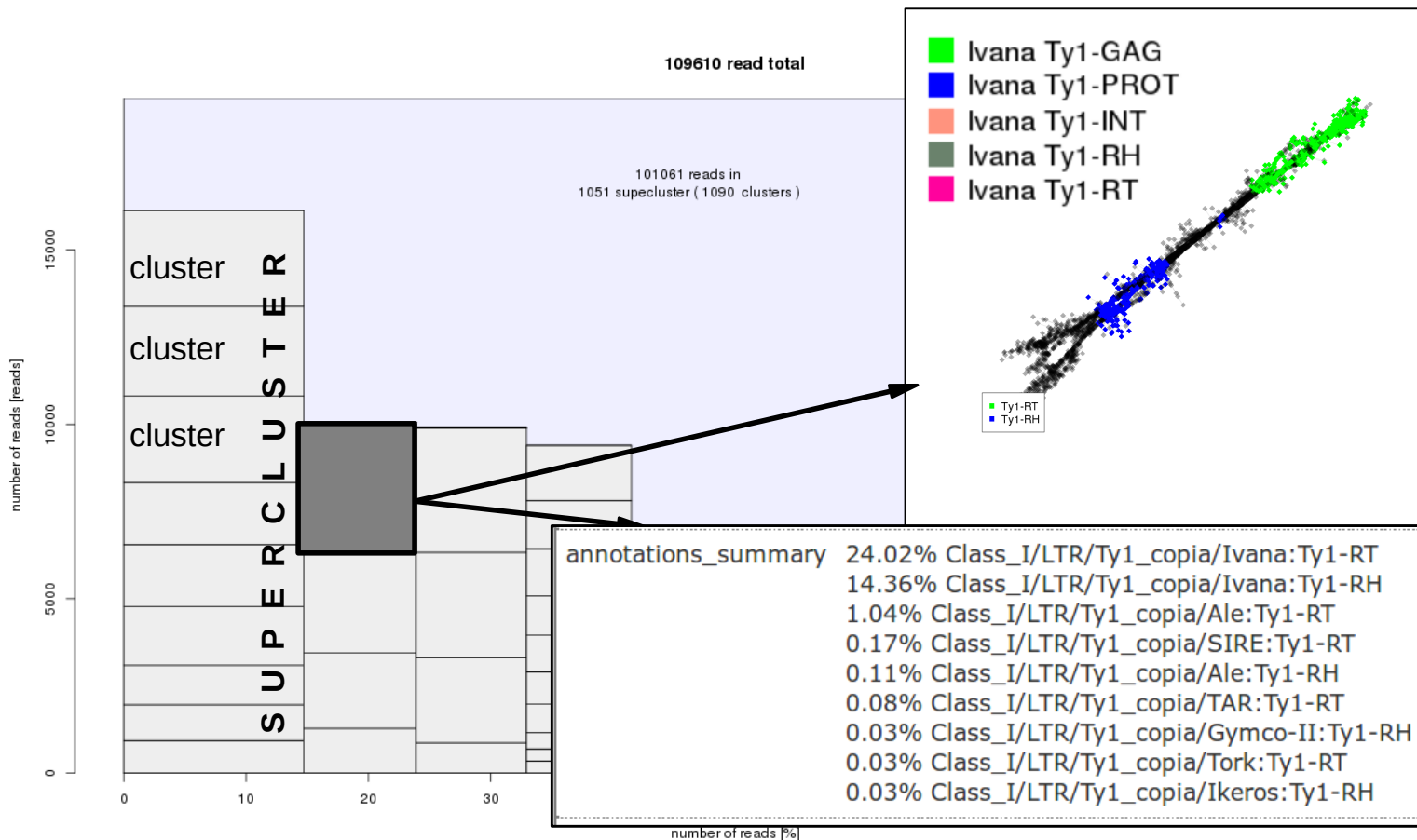
- HTML reports
- Visualization
- Automatic classification

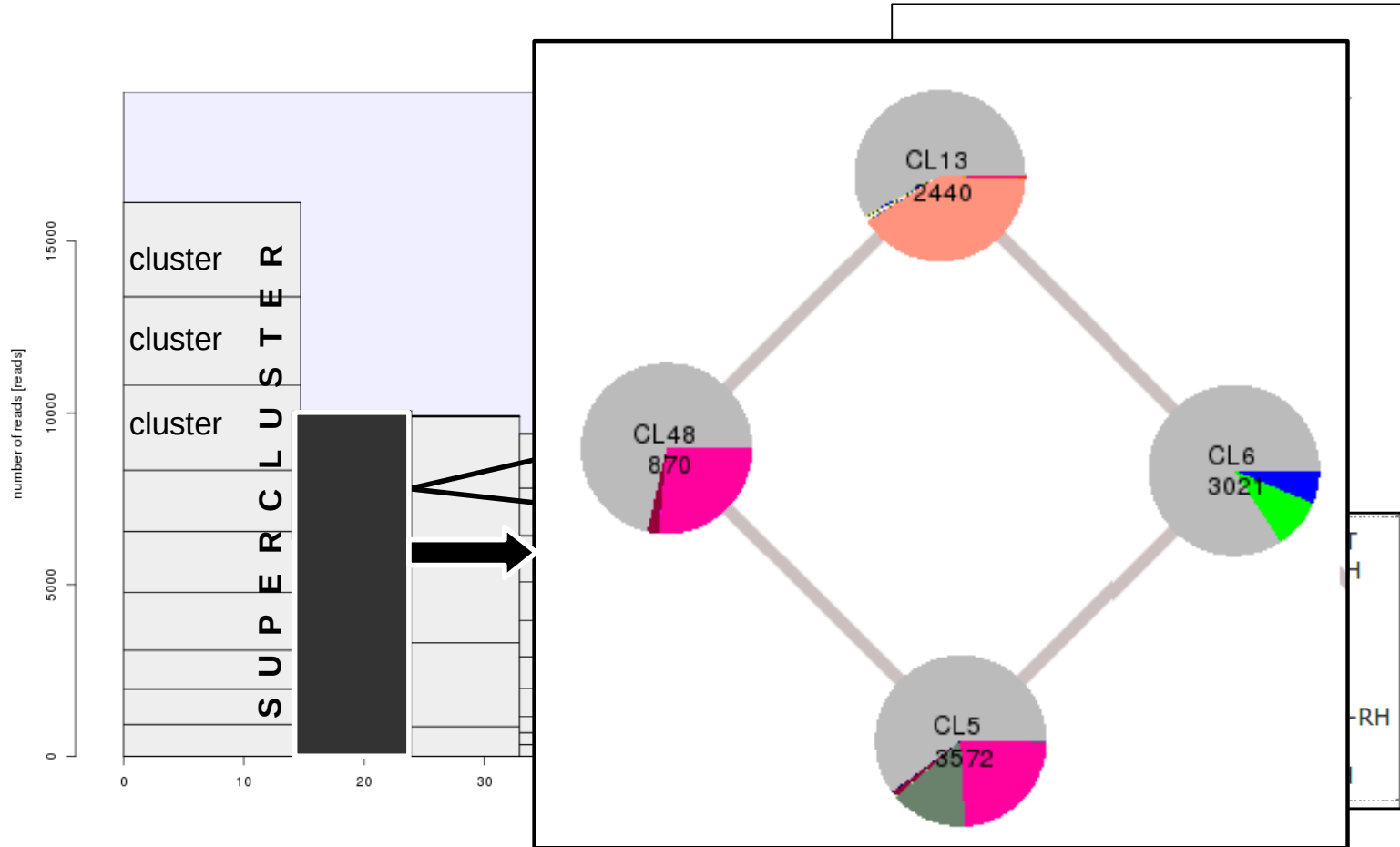


Report

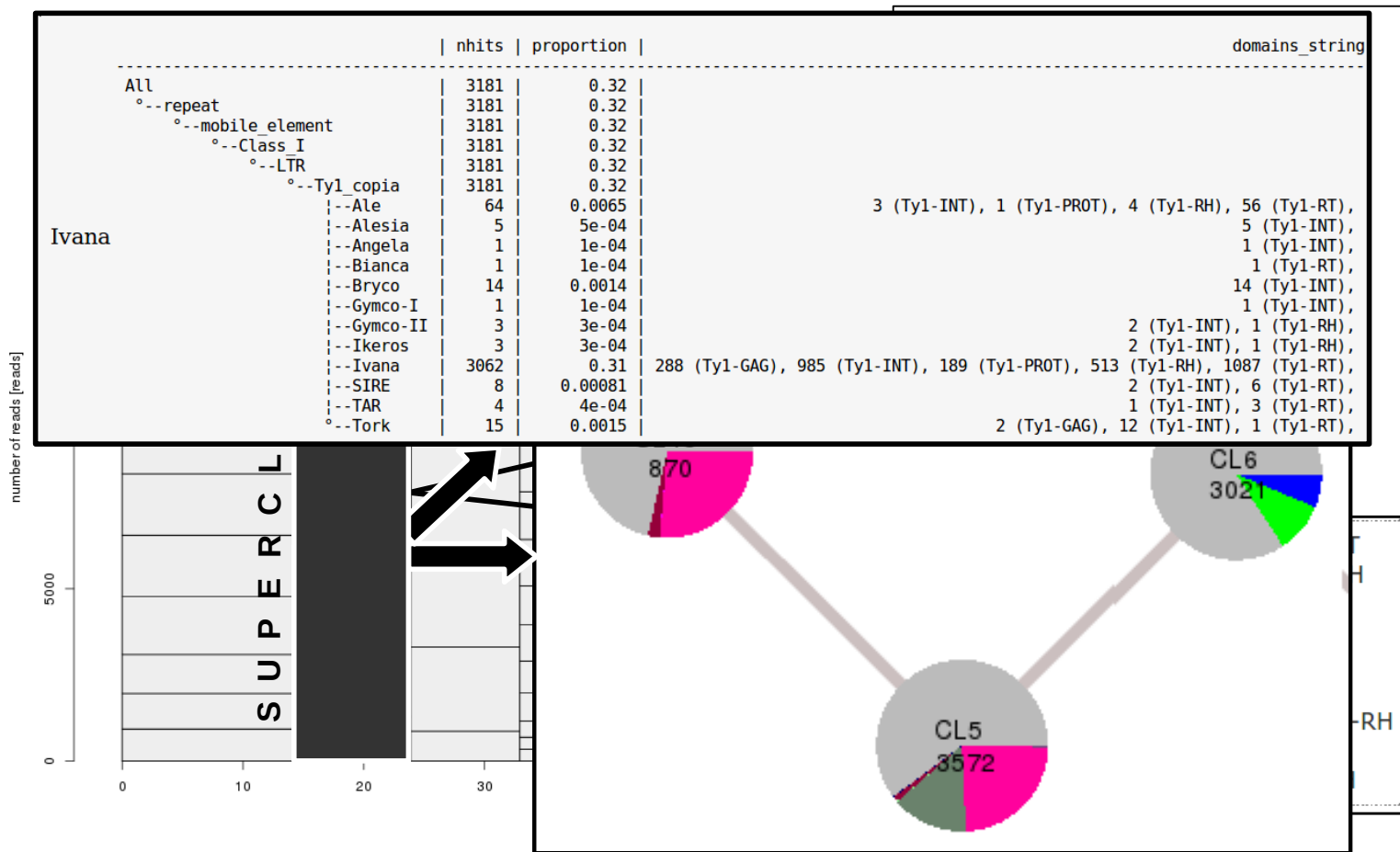


Report

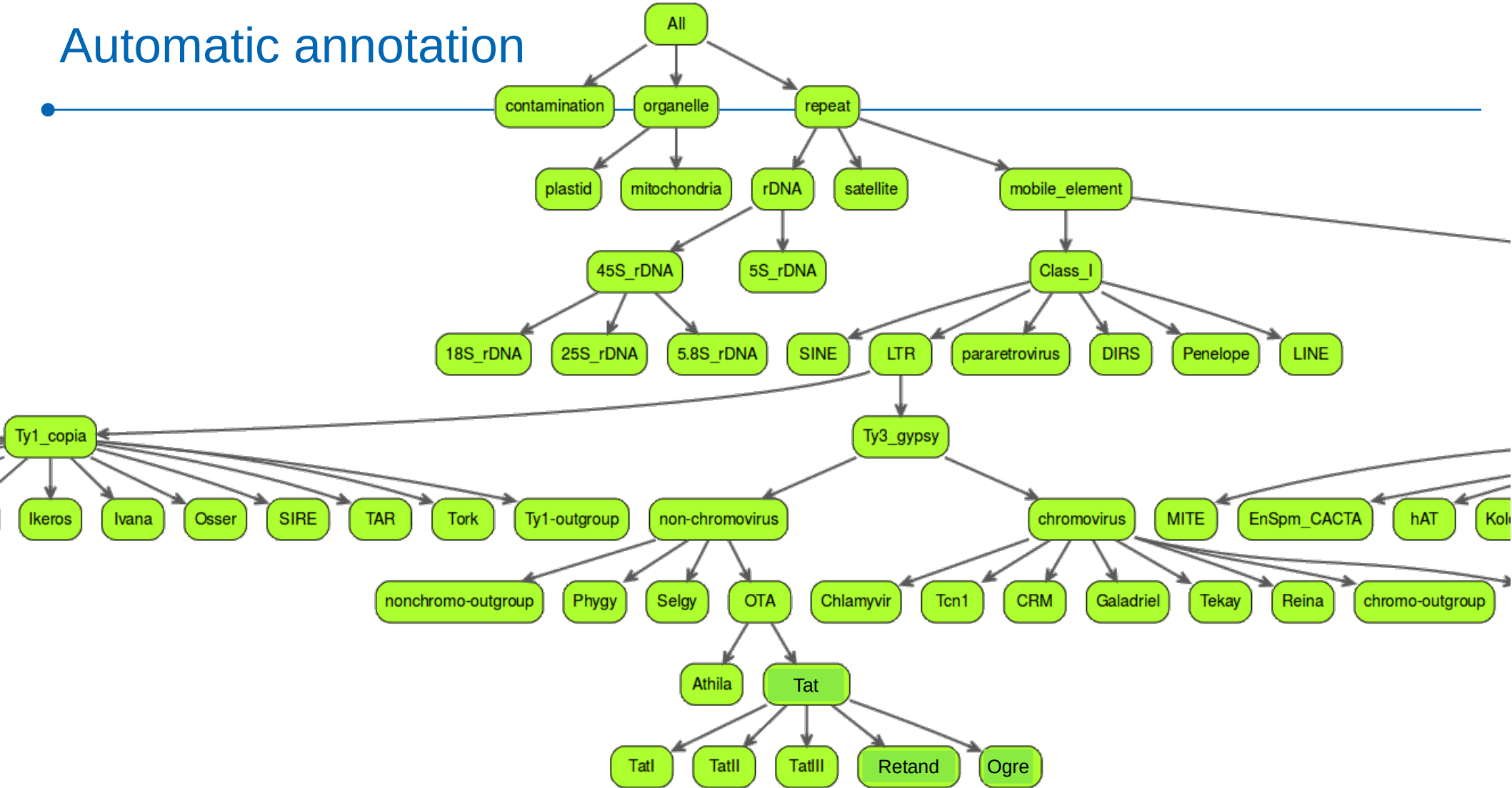




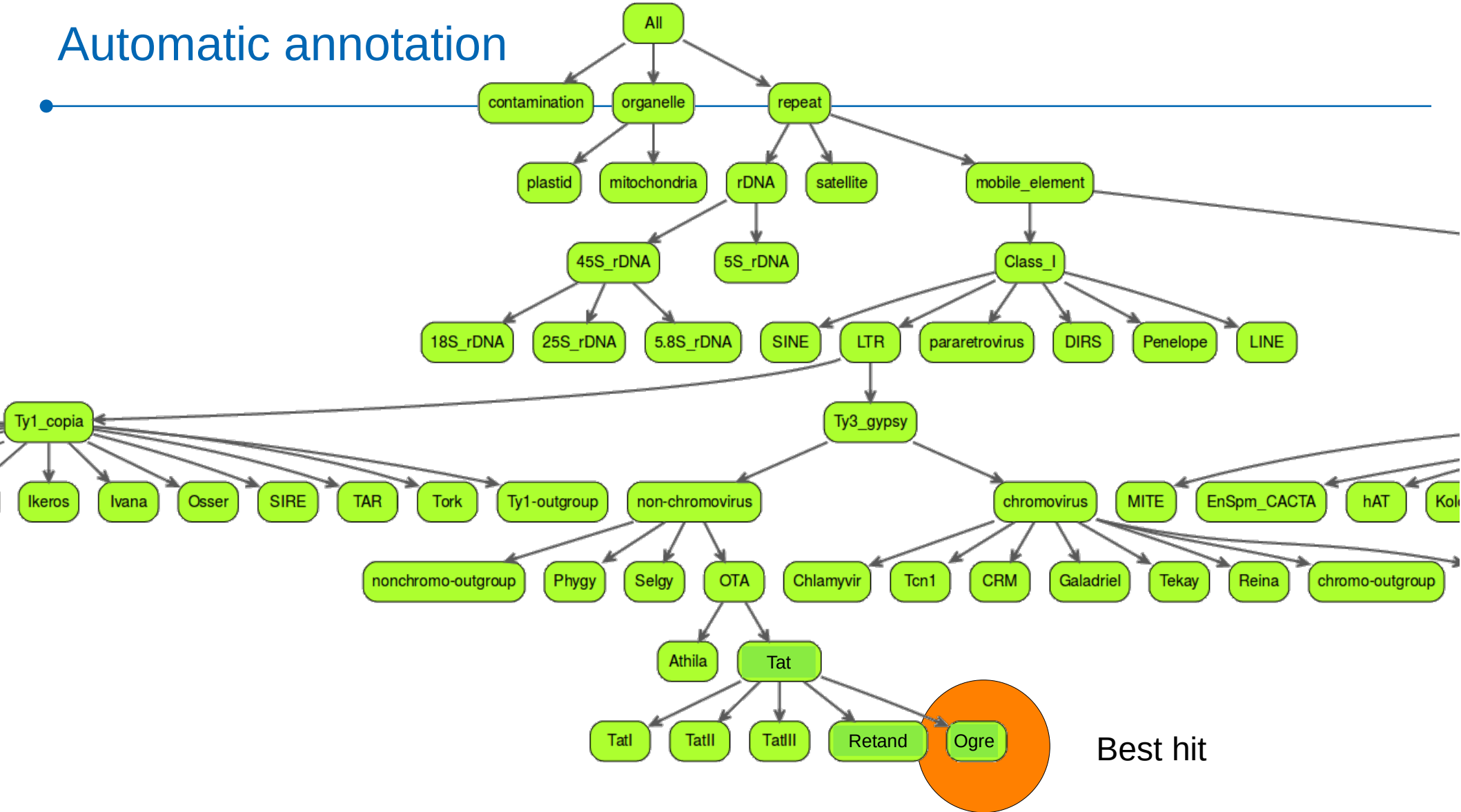
Reporting



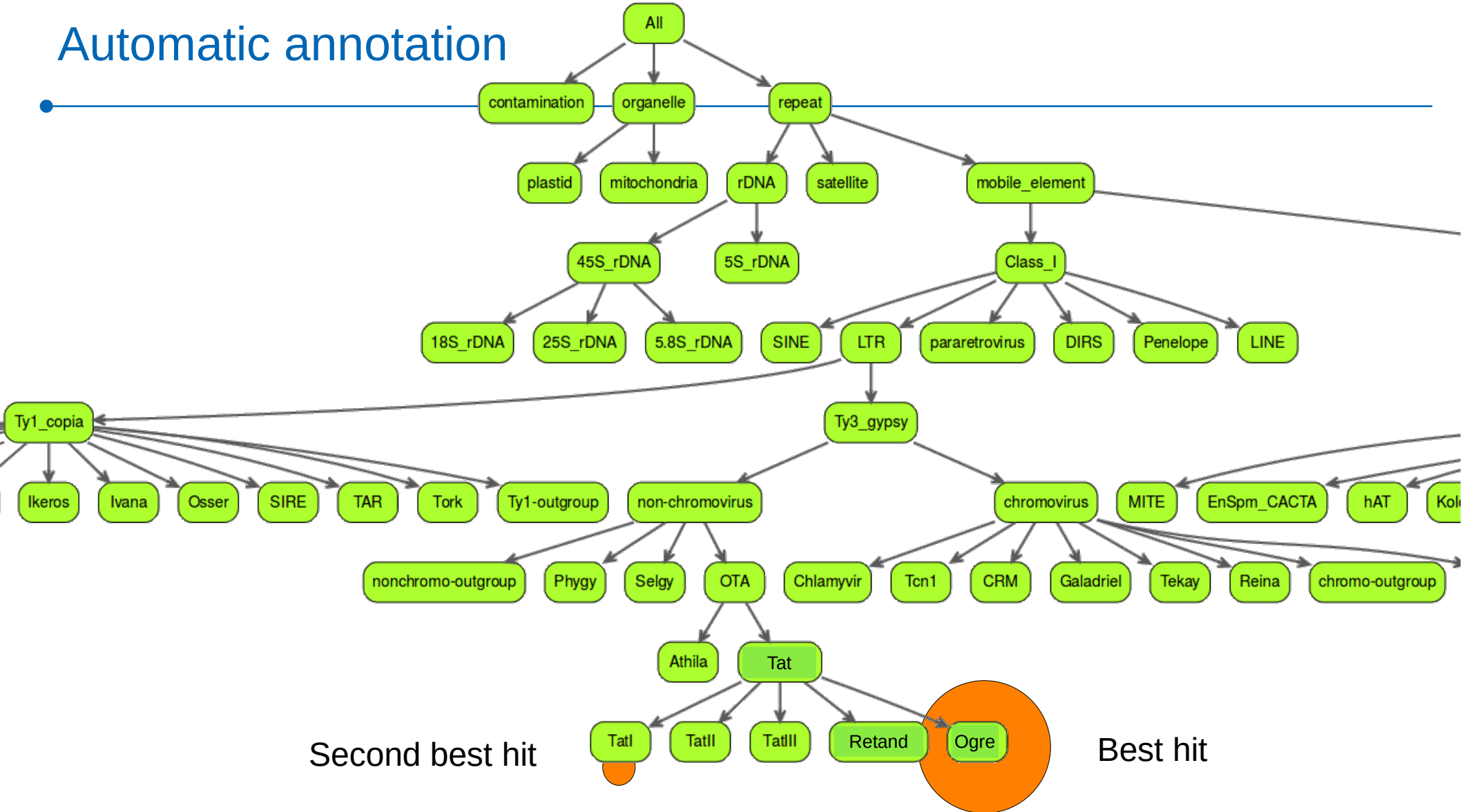
Automatic annotation



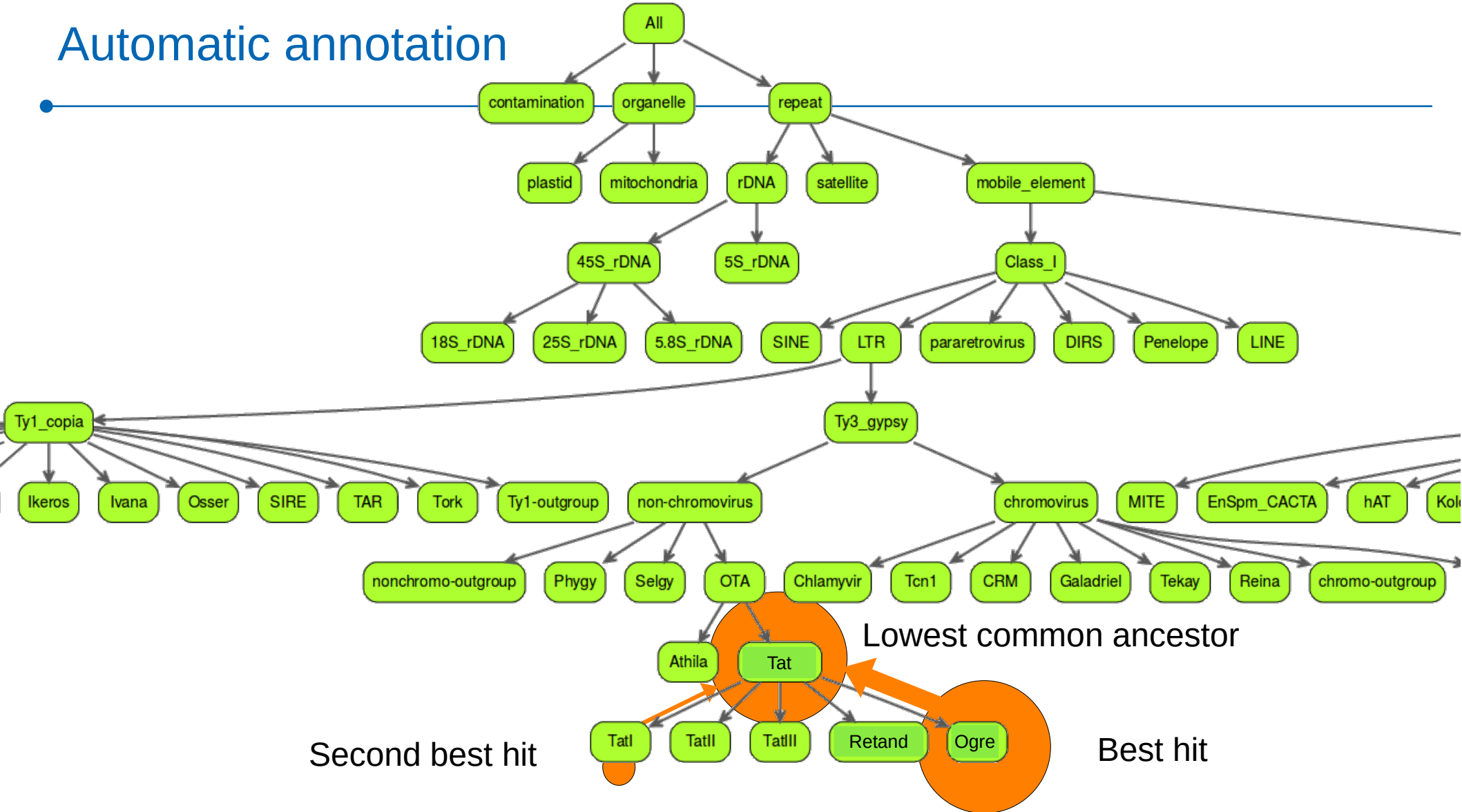
Automatic annotation



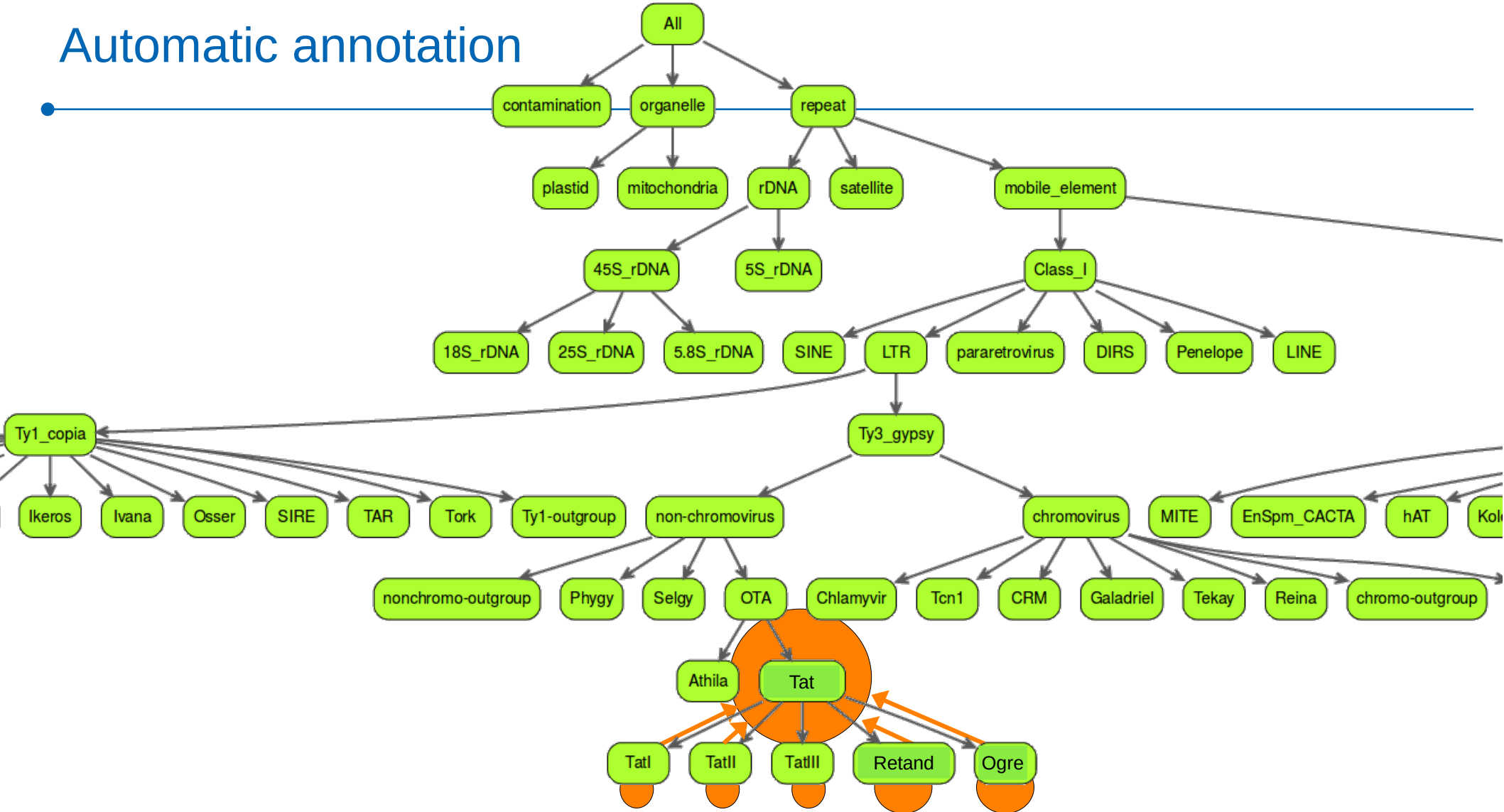
Automatic annotation



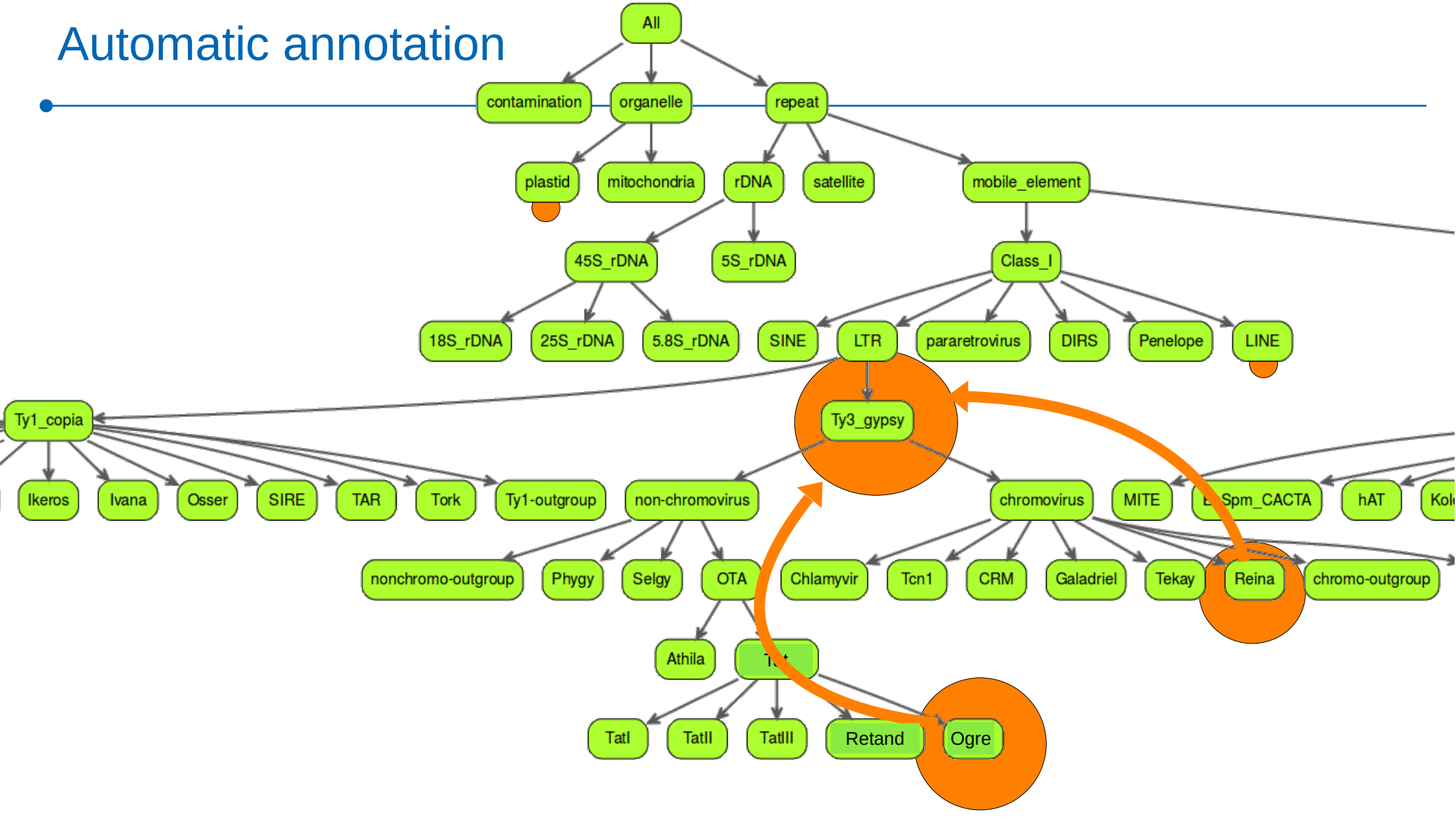
Automatic annotation



Automatic annotation

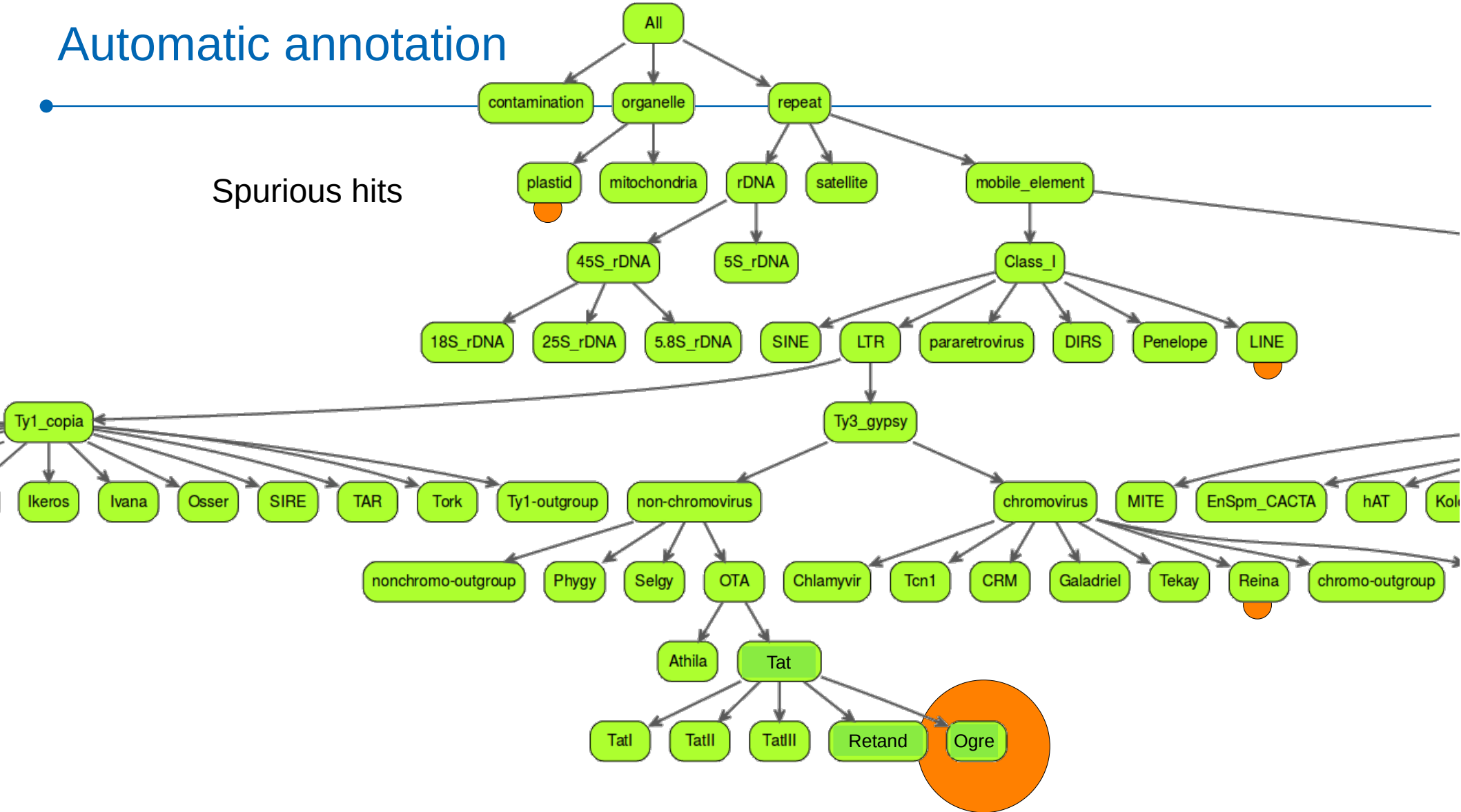


Automatic annotation



Automatic annotation

Spurious hits



Automatic annotation

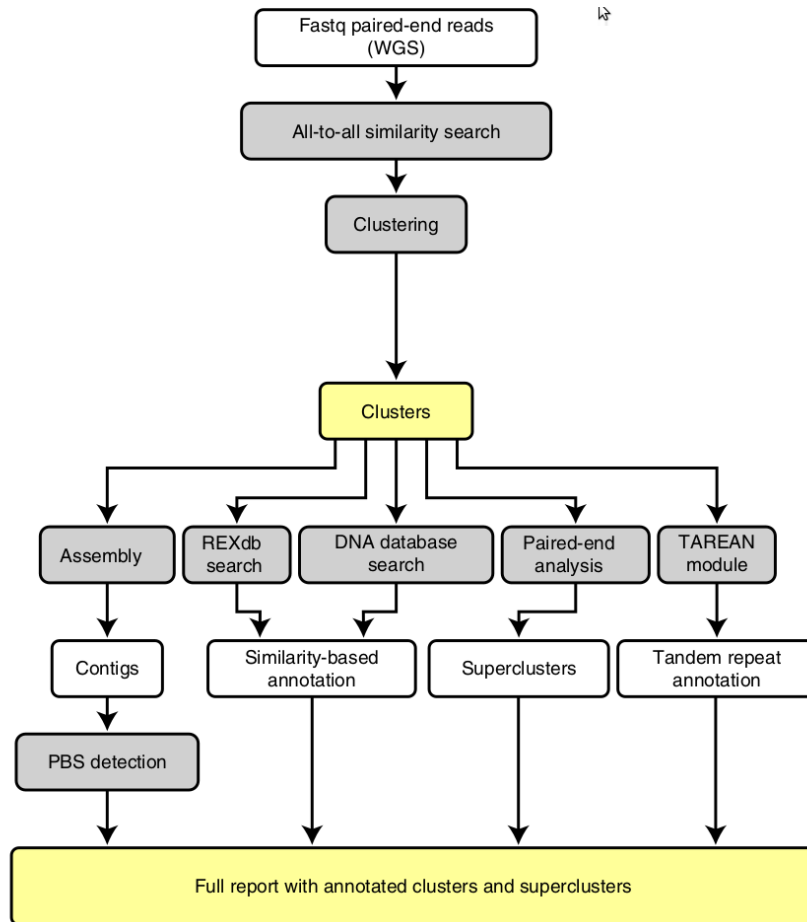
	Proportion[%]	Nsuperclusters	Nclusters	Nreads
Unclassified_repeat (conflicting evidences)	4.06	2	5	67995
--rDNA	0	0	0	0
--45S_rDNA	0.29	2	4	4823
--18S_rDNA	0.04	1	1	653
--25S_rDNA	0.02	1	1	321
--5.8S_rDNA	0	0	0	0
--5S_rDNA	0.12	1	1	1955
--satellite	8.78	33	33	147033
--mobile_element	0	0	0	0
--Class_I	0	0	0	0
--SINE	0	0	0	0
--LTR	0.77	2	5	12931
--Ty1_copia	0	0	0	0
--Ale	0	0	0	0
--Alesia	0	0	0	0
--Angela	0	0	0	0
--Bianca	0.14	1	1	2285
--Bryco	0	0	0	0
--Lyco	0	0	0	0
--Gymco-III	0	0	0	0
--Gymco-I	0	0	0	0
--Gymco-II	0	0	0	0
--Ikeros	0	0	0	0
--Ivana	0.18	2	2	3020
--Gymco-IV	0	0	0	0
--Osser	0	0	0	0
--SIRE	9.57	5	22	160206
--TAR	0.26	5	5	4355
--Tork	0.36	1	1	5947
--Ty1-outgroup	0	0	0	0
--Ty3_gypsy	0	0	0	0
--non-chromovirus	0	0	0	0
--non-chromo-outgroup	0	0	0	0

Automatic annotation

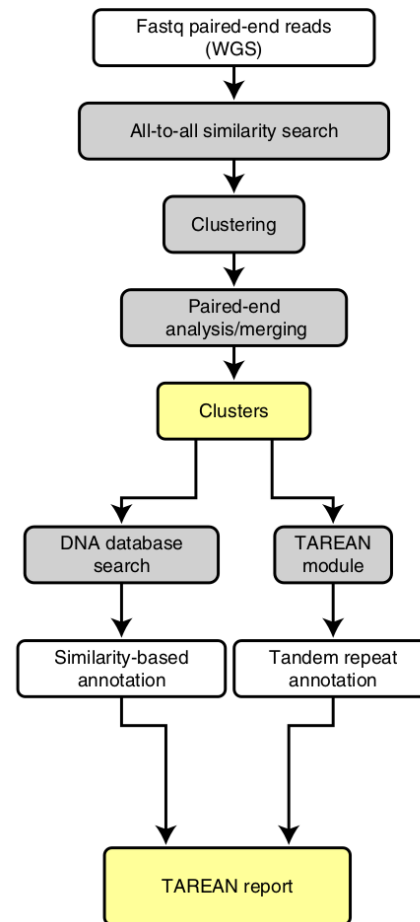
	Proportion[%]	Nsuperclusters	Nclusters	Nreads
Unclassified_repeat (conflicting evidences)	4.06	2	5	67995
--rDNA	0	0	0	0
--45S_rDNA	0.29	2	4	4823
--18S_rDNA	0.04	1	1	653
--25S_rDNA				
--5.8S_rDNA				
--5S_rDNA				
--satellite				
--mobile_element				
--Class_I				
--SINE				
--LTR				
--Ty1_copia				
--Ale				
--Alesia				
--Angela				
--Bianca				
--Bryco				
--Lyco				
--Gymco-III				
--Gymco-I				
--Gymco-II				
--Ikeros				
--Ivana				
--Gymco-IV				
--Osser				
--SIRE				
--TAR				
--Tork				
--Ty1-outgroup				
--Ty3_gypsy				
--non-chromovi				
--non-chro				



RepeaExplorer

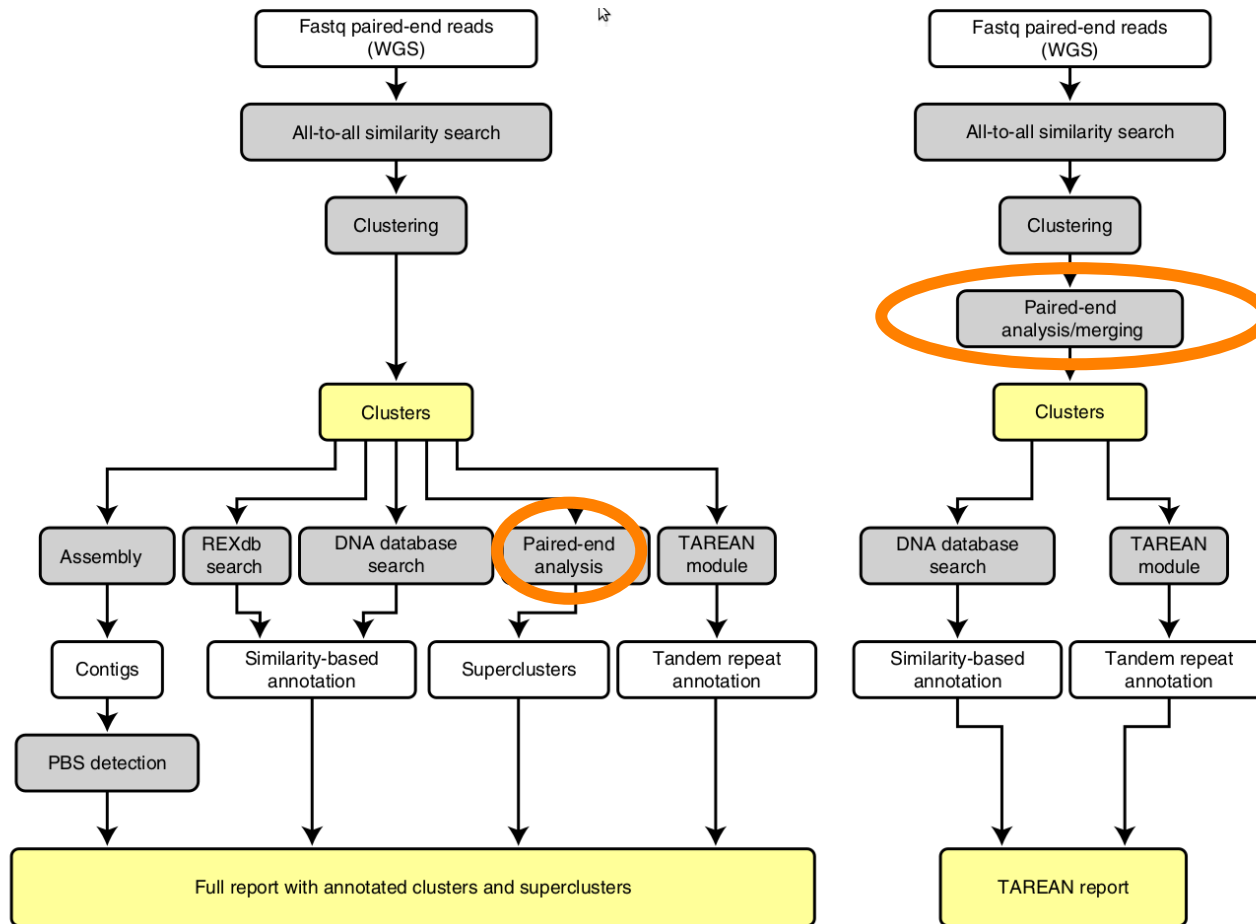


TAREAN



RepeaExplorer

TAREAN



RepeatExplorer Related Tools

- DANTE – **D**omain based **A**nnotation of **T**ransposable **E**lements
 - assembly annotation using REXdb
 - same TE classification system as RepeatExplorer based on REXdb
- Profrep
 - assembly annotaion based on RE results
- ChIP-Seq Mapper
 - Inentification of repeats associated with CENH3 or with a epiginetic marks

Availability

RepeatExplorer Galaxy Server

<https://repeatexplorer-elixir.cerit-sc.cz/>

regalaxy@rt.cesnet.cz

Support:

Martina Macháč
Zdeněk Salvét
Miroslav Ruda
Ivana Křenková



Availability

Command line tools

<https://bitbucket.org/repeatexplorer/>

ChIP-Seq Mapper, RepeatExplorer utilities

https://bitbucket.org/petrnovak/repex_tarean

RepeatExplore with TAREAN

<https://github.com/kavonrtep/dante>

DANTE

<https://github.com/kavonrtep/SeqGrapheR/>

SeqGrapheR

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Thank you!

Questions?